Supplementary Material

Reproductive hormones affect follicular cells and ooplasm of Stage I and II oocytes in zebrafish

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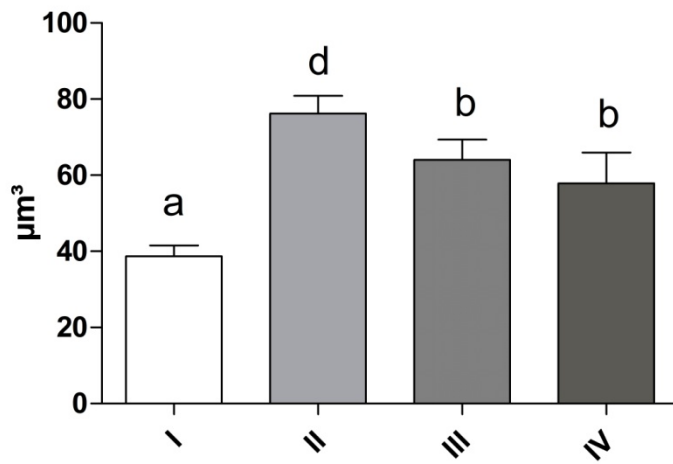
Fig. S1. The volume-weighted nuclear volume of granulosa cells in control follicles from different stages. Data are given as mean ± s.e.m. derived from six individual female zebrafish ($n = 6$). Different letters represent significant differences at $P < 0.05$.
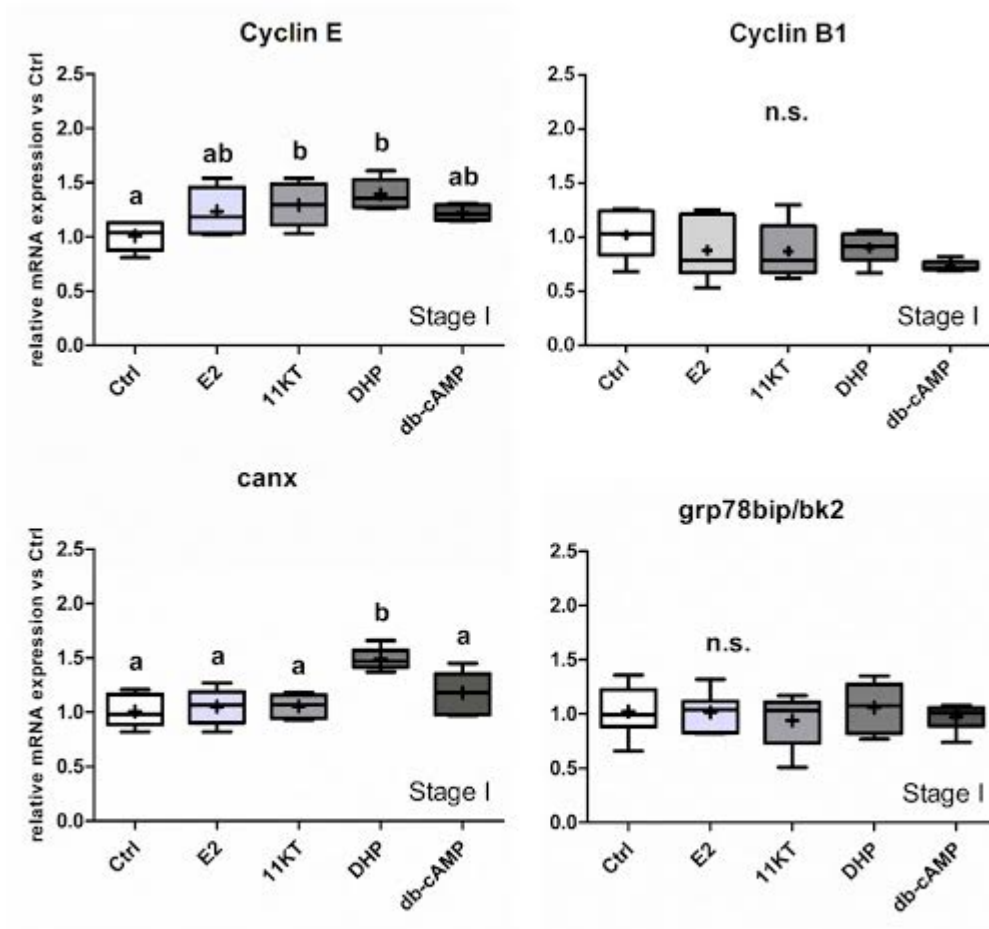
Fig. S2. mRNA expression levels of f cyclin E and cyclin B1 (cell cycle) and of canx and grp78/bip (endoplasmic reticulum resident proteins) at stage I follicles after 48 h of exposure to different hormones; solvent control Ctrl, 17β-estradiol E2, 11 keto-testosterone 11KT, dihydroprogesterone DHP, dibutyryl cyclic AMP db-cAMP. Data are displayed as box-whisker plots derived from 6 individual female zebrafish ($n = 6$). The outer bars cover the 5 and 95% percentiles, the box the 25 and 75% percentiles, the inner bar the median and the cross the mean. Different letters represent significant differences at $P < 0.05$.

Table S1.   Overview of the sequences, amplification efficiencies, and annealing temperature of the primers used for real-time PCR in zebrafish (*Danio rerio*) follicles

Gene name	Forward primer 5' – 3'	Reverse primer 5' – 3'	Amplification efficiency [%]	Annealing temperature [˚C]
cyclin B, *ccnb1*	TGAAGAAGAAGGAGGTGAAGG	CATAGGAACAGGAGAGGAAGG	92.5	55
cyclin E, *ccne*	AAGGAATAGCAGCAGATG	GAAGGAAGTCAAGAGATGG	89.8	55
calnexin, *canx*	TCTGGTGCTCATCATCGTCTTCTG	TGCCTCTGGTTCTTCATCCTTGG	85.5	59
glucose regulated protein 78 KDa, *grp78/bip*	GGACGATAAGAAGGAGAGTG	GTATGACGGAGTGATGCG	95.1	55
elongation factor 1 α, *ef1a*	ATCCGTCGTGGTAATGTGG	TGAGCAGTGTGGCAATCC	92.8	59
ribosomal protein L8, *rpL8*	ATAGTCTGCTGTCTGGAGGAG	TCGGGATTGTGGGAAATAACG	90.6	59