Wildlife Research

Supplementary Material

Chlamydia in wild Australian rodents: a cross-sectional study to inform disease risks for a conservation translocation

Fiona Knox^{A,B,*}, Martina Jelocnik^{C,D}, Nahiid Stephens^A, Colleen Sims^B, Bethany Jackson^A, Saul Cowen^B, Kelly Rayner^B, Sean Garretson^B, Lian Yeap^A, Kristin Warren^A, and Rebecca Vaughan-Higgins^A

^ASchool of Veterinary Medicine, Murdoch University, Murdoch, WA 6150, Australia.

^BBiodiversity and Conservation Science, Department of Biodiversity, Conservation and Attractions, Woodvale, WA 6026, Australia.

^cCentre for Bioinnovation, University of the Sunshine Coast, Sippy Downs, Qld 4556, Australia.

^DSchool of Science, Technology and Engineering, University of the Sunshine Coast, Sippy Downs, Qld 4556, Australia.

^{*}Correspondence to: Fiona Knox School of Veterinary Medicine, Murdoch University, Murdoch, WA 6150 Australia Email: Fiona.knox@murdoch.edu.au



Supplementary Figure S1: Cramer's V coefficient matrix assessing for correlation between categorical variables for all rodent samples.

Results indicate strong and moderate correlation between several categorical variables for the rodent dataset in this study.

Assay	Target gene	Primers and probe	Sequence (5' - 3')	Amplicon size (bp)	Annealing Temperature (°C)	Reference
Chlamudiacoac	23S rRNA	23S-F	CTGAAACCAGTAGCTTATAAGCGGT	111	60	Ehricht et al. (2006)
		23S-R	ACCTCGCCGTTTAACTTAACTCC			
235 FRINA QPCR		23S-Probe	FAM-CTCATCATGCAAAAGGCACGCCG-TAMRA			
<i>C. pecorum</i> qPCR	CpecG_0573	Cpec F3	ATCGGGACCTTCTCATCG	200	57	Jelocnik et al.
		Cpec B3	GCTGTTGTAAGGAAGACTCC	209		(2019)
Partial Chlamydiales	16S rRNA	16S F	GATGAGGCATGCAAGTCGAACG	476	60	Kasimov et al. (2022)
16S rRNA PCR		16S R	GGAGTTAGCCGGTGCTTCTTTAC			

Table S1: Assays and primer sets used in this study for the detection and characterisation of chlamydial DNA.

Table S2: Influence of sampling site on test-prevalence

No significant difference in test prevalence was detected between anatomical sites. Molecular evidence of *Chlamydiaceae* was detected in rodents from all anatomical sites and faecal samples.

Anatomical site								
Pooled Samples	PCR result	Significance test	Individual samples ¹	PCR result	Significance test			
Oral-conjunctival Rectal-faecal:	7/79 10/63;	X ² (2, n=193) = 1.69, <i>P</i> = 0.43	Conjunctival Oral	4/26 2/26	Fisher's Exact Test P =0.94			
Urogenital	7/51		Rectal	4/26				
			Urogenital	3/26				

^J Individual samples were analysed separately from Bernier Island only

Table S3: Histopathology results and tissue *C. muridarum* qPCR results from rodents that had tested positive to *Chlamydiaceae* on Dirk Hartog Island.

One house mouse had histopathological findings that could be consistent with known pathologies of *C. muridarum* in laboratory rodents, demonstrating a subacute suppurative bronchopneumonia and bronchiolitis, but lung tissue from this individual was test negative by *C. muridarum* qPCR. All submitted tissue was test negative by the *C. muridarum* specific qPCR.

Species	Tissue examined	Abnormal tissue findings	Site positive (<i>Chlamydiaceae</i> qPCR)	Tissue subject to <i>C. muridarum</i> PCR
Sandy Inland mouse	Kidney, liver, testis, epididymis, brain, heart, salivary gland and lymph node, lung, adrenal, skin, spleen, oesophagus, stomach, duodenum, pancreas, ileum, colon.	None	Urogenital	Lung
House mouse	Kidney, spleen, lung, uterus, small intestine, liver.	Liver: mild, diffuse, subacute vacuolar hepatopathy (consistent with glycogen).	Rectal	Faeces
House mouse	Brain, liver, heart, skeletal muscle, stomach, salivary gland and lymph node, adrenal, skin, spleen, jejunum, caecum, colon, kidney.	Kidney – marginal, multifocal, chronic lymphoplasmacytic interstitial nephritis.	Rectal and urogenital	Gastrointestinal (GIT) sections (caecum, stomach, small intestine, colon) Lung
House mouse	Salivary gland and lymph node, skeletal muscle, kidney, brain, testis and epididymis, pancreas, skin, lung, small intestine, spleen, heart, liver, caecum.	None	Rectal	GIT sections (caecum, stomach, small intestine, colon) Lung
House mouse	Heart, salivary gland and lymph node, brain, liver, kidney, stomach, skeletal muscle, skin, spleen, lung, colon, small intestine.	Mandibular lymph node lymphofollicular hyperplasia.	Rectal	GIT sections (caecum, stomach, small intestine, colon) Lung
House mouse	Heart, salivary gland and lymph node testis, brain, liver, kidney, skeletal muscle, stomach, skin, lung, pancreas, spleen, ileum, jejunum.	Mandibular lymph node - medullary histiocytosis (drainage reaction).	Conjunctival, oral and urogenital	GIT sections (caecum, stomach, small intestine, colon) Lung
House mouse	Lung, caecum, salivary gland and lymph node, kidney, brain, liver, skin, mammary gland, skeletal muscle, pancreas, spleen, small intestine, cervix, uterine horn, ovary, adrenal, colon, heart, stomach.	Lung – moderate, focally extensive, subacute suppurative bronchopneumonia and bronchiolitis. Caecum – putative oxyuriasis. Mandibular lymph node – medullary histiocytosis (drainage reaction). Kidneys – marginal, focal and bilateral, chronic lymphoplasmacytic interstitial nephritis.	Urogenital	GIT sections (caecum, stomach, small intestine, colon) Lung