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Additions to *Amanita* (Amanitaceae, Agaricales) section *Arenariae* from south-western Australia

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Abstract. A recent molecular phylogeny of *Amanita* recognises three subgenera and 11 sections. Members of subgenus *Amanitina* are characterised by amyloid spores and a mycorrhizal habit. Section *Arenariae* falls within this subgenus. Members of this section are known only from southern Australia; they are either sequestrate (secotioid) or agaricoid and lack clamp connections. We describe the following three additional secotioid species: *Amanita arenarioides* Bougher, E.M.Davison & Giustiniano, *A. compacta* Bougher, E.M.Davison & Giustiniano and *A. pseudoarenaria* E.M.Davison, Giustiniano & Bougher, which are separated on macroscopic appearance, spore shape and genetic sequences. We also describe two agaricoid species, namely, *A. pupatuju* E.M.Davison, Giustiniano, McGurk & E.L.J.Watkin, and *A sabulosa* E.M.Davison & Giustiniano, which are separated on bulb shape and genetic sequences. We provide expanded descriptions of *A. arenaria* (O.K.Mill. & E.Horak) Justo and *A. griselloides* D.A.Reid; we also synonymise *A. dumosorum* D.A.Reid with *A. peltigera* D.A.Reid. A revised diagnosis and description of section *Arenariae* is provided, together with a key to currently recognised member of this section.

Keywords: Basidiomycota, multi-gene phylogenetics, taxonomy, ITS variation.

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Introduction

Amanita Pers. is the largest genus in Amanitaceae, with ~1000 described species worldwide; however, only ~600 are well defined (Yang et al. 2018). This genus is important ecologically because most species are mycorrhizal with woody plants. It is economically important because some species are comestible, and others are poisonous (Cui et al. 2018). The genus is defined by the field character of a membranous or friable universal veil, and by the microscopic and ontogenetic characters of bilateral lamella trama, acrophysalidic stipe trama and schizohymenial development (Bas 1969, pp. 299-333). Like many mushrooms, *Amanita* spp. are phenotypically variable; consequently, many species cannot be confidently identified using field characters alone; microscopic examination is also essential. Molecular analyses too have been instrumental in both identifying new species and clarifying relationships within the genus (Cui et al. 2018; Yang et al. 2018).

Within Australia, agaricoid species have been monographed by Reid (1980) and Wood (1997). Additional descriptions and clarifications have been made by Miller

(1991, 1992), Grgurinovic (1997), Davison (2011), Davison and Giustiniano (2020), Davison *et al.* (2013, 2015, 2017*a*, 2017*b*, 2020) and McGurk *et al.* (2016). In addition, there are sequestrate species that were assigned to *Amarrendia* Bougher & T.Lebel and *Torrendia* Bres., genera that are now synonymised with *Amanita* (Miller and Horak 1992; Bougher 1999; Bougher and Lebel 2002; Justo *et al.* 2010). There are ~100 named Australian species, but many more await description.

The family Amanitaceae has been recently revised by Cui et al. (2018), using concatenated datasets of nuLSU (28S nuclear ribosomal large subunit rRNA gene), β -tubulin, efl- α (elongation factor 1- α), and rpb2 (RNA polymerase II) gene regions. Within Amanita, they recognise three subgenera, namely, Amanita, Amanitina (E.-J.Gilbert) E.-J. Gilbert and Lepidella Beauseign. Most Australian species fall within subgenus Amanitina because they have amyloid spores, a bulb at the base of the stipe, and are likely to be mycorrhizal. Cui et al. (2018) recognised the following six sections within Amanitina: section Amidella (E.-J.Gilbert) Konrad & Maubl., section Arenariae Zhu L.Yang, Y.Y.Cui & Q.Cai, section

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Phalloideae (Fr.) Quél., section Roanokenses Singer, section Strobiliformes Singer ex Zhu L.Yang, Y.Y.Cui & Q.Cai, and section Validae (Fr.) Quél. These sections can be recognised morphologically by differences in the pileus margin, form of the universal veil, and presence or absence of clamp connections.

Members of section Arenariae are agaricoid or sequestrate, their basidia are thin- or thick-walled and lack clamp connections; agaricoid species have an appendiculate pileus margin and remnants of universal veil on the pileus (Cui et al. 2018, pp. 106, 117). Five species have been described; they are known only from southern Australia. The type is A. arenaria (O.K.Mill. & E.Horak) Justo, a sequestrate (secotioid) species from the south-west of Western Australia (WA). Amanita arenaria is unusual because it has inamyloid spores. whereas they are amyloid in agaricoid members of this section. There are a further five sequestrate species of Amanita known from Australia, and they cluster together in section Amarrendiae (Bougher & Lebel) Zhu L.Yang, Y.Y. Cui, Q.Cai & L.P.Tang, which are in subgenus Amanita, not subgenus Amanitina (Justo et al. 2010; Cui et al. 2018, pp. 19, 44).

Miller and Horak (1992) described A. arenaria (as Torrendia arenaria O.K.Mill. & E.Horak) from collections they had made while visiting WA in 1985, 1989 and 1991. Miller split two of his collections (OKM 24013, VPI 679; OKM 24018, VPI 412) and deposited duplicates in the Western Australian Herbarium (PERTH). Within T. arenaria, they described T. arenaria f. lutescens O.K.Mill. & E.Horak for material showing yellow stains on the stipe and pileus or slowly turning yellowish on bruising or cutting. Justo et al. (2010) re-examined 10 of Miller's collections of T. arenaria, together with other Torrendia species, and used the nuLSU gene region to determine whether they were closely related to Amanita, as had been suggested by Bas (1975). They showed all Torrendia species nested within Amanita and transferred all species to Amanita. Their results showed that these secotioid species were not monophyletic because they fell into three well-supported clades in different parts of the genus. They also showed that four of the collections identified by Miller and Horak (1992) as T. arenaria were an undescribed species that was morphologically and molecularly distinct, and described it as A. pseudoinculta Justo. One of the collections examined by Justo et al. (2010) was T. arenaria f. lutescens; however, the nuLSU sequence from this collection did not differ from sequences from collections identified as T. arenaria f. arenaria. They also noted that the six A. arenaria collections consistently clustered into two subgroups that differed in spore shape; those from the type locality were ellipsoid to elongate (Q = 1.55-1.72), whereas spores of the others were broadly ellipsoid to ellipsoid (Q = 1.28-1.36). They suggested that this either represented two cryptic species, or there was high genetic divergence among collections from different localities.

While reviewing all *A. arenaria* collections in PERTH and in private collections, we have noted there are at least three species that are macroscopically similar but can be separated by spore shape and molecular sequences. We describe these in this paper, together with another secotioid species that differs macroscopically from *A. arenaria*.

There are four agaricoid species that are recognised as being members of section *Arenariae*; these are *A. lesueurii* E. M.Davison, *A. peltigera* D.A.Reid, *A. wadjukiorum* E.M. Davison and *A. wadulawitu* McGurk, E.M.Davison & E.L.J. Watkin (Cui *et al.* 2018; Davison *et al.* 2020). In this paper, we describe two additional species. Many more described species may be recognised as being members of this section if widespread sequencing of described Australian amanitas is conducted.

Reid (1980) described *A. griselloides* D.A.Reid as having a layered universal veil on the pileus; however, examination of the type by E. M. Davison found that the superficial layer was formed by hyphomycetes. We provide a revised description in this paper.

The type of A. peltigera was described from South Australia (SA) as having a saccate volva and lacking a partial veil (Reid 1980; Davison et al. 2020). However, examination of collections from SA and the south-west of WA showed that they form a single clade exhibiting variation in both the persistence of the partial veil and remains of the universal veil at the top of the bulb (Davison et al. 2020). Reid (1980) also described A. dumosorum D.A.Reid, which he considered to be similar to A. peltigera apart from having a partial veil, lacking a saccate volva, and lacking infrequent large, inflated cells in the universal veil on the pileus. However, examination of the type of A. dumosorum (K(M) 236387) by E. M. Davison found infrequent large, inflated cells in this tissue. The other differences noted by Reid (1980) occurred in collections of A. peltigera (Davison et al. 2020); because these two species cannot be separated by their field characters or micromorphology, they are synonymised in this paper.

The nuclear ribosomal internal transcribed spacer (ITS) region has been used as a barcode marker for species discrimination within the Basidiomycota (Schoch et al. 2012). Hughes et al. (2013) found less than 2% base pair divergence among haplotypes within an individual, and Cai et al. (2014) used ITS sequences to separate Amanita species within section *Phalloideae*. The *ITS* region does not appear as useful among Australian species. For example, Davison et al. (2017a) found that ITS sequences fail to separate three species from section *Phalloideae*, which differ in spore shape and are geographically separated, and within A. peltigera, the base pair divergence among haplotypes from the same individual is between 0.2 and 5.8% (Davison et al. 2020). In this paper, we determine the difference between ITS clones from both secotioid and agaricoid members of section Arenariae, to determine whether they show divergence similar to that observed in A. peltigera.

This paper has four aims. First, molecular sequences are used to check the placement of the undescribed species and *A. griselloides* D.A.Reid within section *Arenariae*, their relationships with one another, and the variation within the *ITS* region of haplotypes from the same individual and from different collections of the same species. Second, we describe three new secotioid and two new agaricoid species, together with a revised description of *A. arenaria* and an expanded description of *A. griselloides*. Third, we synonymise *A. dumosorum* with *A. peltigera*. Last, we provide revised

diagnosis and description of section Arenariae and a key to currently recognised members of this section.

Materials and methods

Taxonomy

The methodology used for describing the macroscopic and microscopic characters largely follows Tulloss (2000). Colour names, including the colour of spores in deposit and other shades of white to cream (designated by the letters A–G), follow Royal Botanic Garden, Edinburgh (1969), whereas colour codes are from Kornerup and Wanscher (1983). In the descriptions of basidiospores (and basidia) the notation [x/y/z] denotes x basidiospores measured from y basidiomes from z collections. Biometric variables for spores follow Tulloss (2000), that is:

- L = the average spore length computed for one specimen examined and the range of such averages
- L' = average spore length computed for all spores measured
- W = the average spore width computed for one specimen examined and the range of such averages
- W' = average spore width computed for all spores measured

- Q = the length:breadth for a single spore and the range of the ratio of length:breadth for all spores measured
- **Q** = the average value of Q computed for one specimen examined and the range of such averages
- Q' = the average value of Q computed for all spores measured

Author citations follow Index Fungorum (see http://www.indexfungorum.org/Names/Names.asp, accessed 19 February 2021). Herbarium codes follow Index Herbariorum (see http://sweetgum.nybg.org/science/ih/, accessed 1 February 2021).

Phylogenetics

DNA extraction, amplification and cloning of the *ITS*, amplification of the *nuLSU*, $efl-\alpha$, rpb2 and β -tubulin regions follow the methodology in Davison et al. (2017b). Sequence data were assembled with Geneious (ver. 10.0.5, www.geneious. com; Kearse et al. 2012). Additional sequences were accessed from GenBank (see http://www.ncbi.nlm.nih.gov/, accessed 2 February 2021; Tables 1, 2).

Any polymerase chain reaction (PCR) chimeras in *ITS* sequences were detected using the 'Remove chimeric reads' function in Geneious, using the UCHIME program (yer. 11.0.667, see https://www.drive5.com/usearch/manual/

Table 1. Voucher information and GenBank accession numbers for *nuLSU* sequences from type of sections and other relevant collections used to place new species in subgenus *Amanitina*

Newly published sequences are shown in bold. nuLSU, nuclear ribosomal large-subunit rRNA region; WA, Western Australia; SA, South Australia

Section	Amanita spp.	Voucher	Locality	nuLSU
Amidella	A. volvata (Peck) Lloyd (type of section)	KA12-1367	Gyeongbuk, South Korea	KF245907
	A. brunneomaculata Zhu L.Yang, Y.Y.Cui & Q.Cai	HKAS70032	Yunnan, China	MH486411
	A. lanigera Y.Y.Cui, Q.Cai & Zhu L.Yang	HKAS89030	Yunnan, China	MH486621
	A. parvicurta Y.Y.Cui, Q.Cai & Zhu L.Yang	HKAS101215	Yunnan, China	MH486745
Arenariae	A. arenaria holotype (type of section)	VPI 679	City of Albany, WA, Australia	GQ925382
	A. arenaria	VPI 412	City of Albany, WA, Australia	GQ925383
	A. arenaria	PERTH 09316213	City of Albany, WA, Australia	MW793397
	A. arenaria	PERTH 07367813	Shire of Denmark, WA, Australia	MW775281
	A. peltigera	AD 282185	Kangaroo Island, SA, Australia	MN900628
	A. peltigera	PERTH 08793514	Shire of Manjimup, WA, Australia	MN900625
	A. peltigera	PERTH 08793581	Shire of Serpentine-Jarrahdale, WA, Australia	MN900627
	A. peltigera	PERTH 09138560	Shire of Mundaring, WA, Australia	MN900626
	A. wadulawitu	PERTH 09144382	Shire of Esperance, WA, Australia	MN918101
	A. wadulawitu	PERTH 09144404	City of Melville, WA, Australia	MN918100
	A. wadulawitu	PERTH 09144390	Shire of Serpentine–Jarrahdale, WA, Australia	MN918098
Phalloideae	A. phalloides (Vaill. ex Fr.) Link (type of section)	HKAS75773	China	JX998060
	A. djarilmari E.M.Davison	PERTH 08776067	Shire of Cuballing, WA, Australia	KY977704
	A. marmorata (Cleland & EJ. Gilbert) EJ.Gilbert	PERTH 08690596	Shire of Denmark, WA, Australia	KY977711
Roanokenses	A. roanokensis Coker (type of section)	FLAS-F-60892	Florida, USA	MH620252
	A. carneiphylla O.K.Mill.	PERTH 08793530	City of Melville, WA, Australia	MN911351

(continued next page)

Table 1. (continued)

Section	Amanita spp.	Voucher	Locality	nuLSU
	A. hiltonii D.A.Reid	PERTH 09004564	Shire of Mundaring, WA, Australia	MT364456
	A. ochrophylla (Cooke & Massee) Cleland	AD-C 49440	SA, Australia	HQ539717
	A. preissii (Fr.) Sacc.	PERTH 08690766	Kings Park, WA, Australia	KY290654
Strobiliformes	A. strobiliformis (Paulet ex Vittad.) Bertill. (type of section)	MB-001177	Germany	MH486895
	A. cinereopannosa Bas	RET 318-8	Maine, USA	HQ539678
	A. cinereoradicata Y.Y.Cui, Q. Cai & ZhuL.Yang	HKAS63641	Yunnan, China	MH486452
Validae	A. excelsa (Fr.) Bertill. (type of section)	HKAS96169	Austria	MH486492
	A. flavoconia G.FAtk.	BW_PH22	Massachusetts, USA	HQ539693
	A. citrina Pers.	BW JLR 102106-1	New Jersey, USA	HQ539679
Amanita	A. subglobosa Zhu L.Yang (outgroup)	HKAS58837	China	JN941152
	A. arenarioides sp. nov. holotype	PERTH 07627025	City of Wanneroo WA, Australia	MW775283
	A. arenarioides	PERTH 07626959	Shire of Gingin, WA, Australia	MW775282
	A. compacta sp. nov. holotype	PERTH 08700524	Shire of Wongan–Ballidu, WA, Australia	MW775285
	A. griselloides	PERTH 09004831	City of Albany, WA, Australia	MW775286
	A. pseudoarenaria sp. nov. holotype	PERTH 08606765	Shire of Manjimup, WA, Australia	MW775284
	A. pseudoarenaria [as A. arenaria]	VPI 551	Kalamunda, WA, Australia	GQ925381
	A. pseudoarenaria [as A. arenaria]	VPI 365	Kalamunda, WA, Australia	GQ925379
	A. pseudoarenaria [as A. arenaria]	VPI 363	Julimar, WA, Australia	GQ925384
	A. pseudoarenaria [as A. arenaria f. lutescens]	VPI 364	Kalamunda, WA, Australia	GQ925380
	A. pseudoarenaria	PERTH 09316183	Shire of Serpentine–Jarrahdale, WA, Australia	MW793396
	A. pseudoarenaria	PERTH 09316191	Shire of Mundaring, WA, Australia	MW793398
	A. pupatju sp. nov. holotype	PERTH 09269525	City of Melville, WA, Australia	MW775287
	A. sabulosa sp. nov. holotype	PERTH 09178759	City of Melville, WA, Australia	MW775279
	A. sabulosa	PERTH 09178775	City of Melville, WA, Australia	MW775280
	A. sabulosa [as A. subalbida]	AD-C54481	Monarto Conservation Park, SA, Australia	HQ539745

uchime_algo.html; Edgar *et al.* 2011). Default settings were used (minimum reporting score 0.3, weight no vote 8, minimum divergence ratio 0.5). The recommended database for chimera detection in fungal *ITS* sequences was selected as the reference database (Abarenkov *et al.* 2020).

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Maximum-likelihood phylogenetic trees were built using MEGA (ver. 5, see https://www.megasoftware.net; Tamura et al. 2011). The best model for each dataset was determined using the Model Function in MEGA. The general time-reversible model (Tavaré 1986) with gamma distribution rates was used to determine the placement of the undescribed species in subgenus Amanitina by using the nuLSU gene region. The Kimura 2-parameter model (Kimura 1980) with invariant sites was used for the $efl-\alpha$ and β -tubulin gene regions. The Kimura 2-parameter model (Kimura 1980) with gamma distribution rates was used for the efl gene regions. The Tamura–Nei model (Tamura and Nei 1993) with invariant sites and gamma distribution rates was

used for concatenated nuLSU, rpb2, $ef1-\alpha$ and β -tubulin gene regions. The Tamura 3-parameter model with gamma distribution was used for the ITS gene regions for the secotioid species (Tamura 1992). Bootstrap consensus trees were inferred from 500 replicates.

Results

Phylogenetics

Placement of undescribed species in subgenus Amanitina

The *nuLSU* is the only gene region available from GenBank for all type species of sections within subgenus *Amanitina*. These, together with additional species (Table 1), show that *A. arenarioides* sp. nov., *A. compacta* sp. nov., *A. griselloides*, *A. pseudoarenaria* sp. nov., *A. pupatju* sp. nov. and *A. sabulosa* sp. nov. form clades within section *Arenariae* with good support (Fig. 1). There are two clades within this section. One well-supported clade contains the secotioid

Table 2. Voucher information and GenBank accession numbers for sequences of species from section Arenariae

Newly published sequences are shown in bold. ITS, nuclear ribosomal transcribed-spacer region; eff-α, translation elongation-factor 1-α region; rpb2, RNA polymerase-II region; B-tubulin, β-tubulin region; WA, Western Australia

	V oucher number	Locality	GenBank numbers ITS	efI - α	rpb2	B-tubulin
A. arenaria PE	VPI 679 PERTH 09316213	City of Albany, WA, Australia City of Albany, WA, Australia	GQ925393 MW795711–MW795714	MW820649	MW820674	MW820671
A. arenaria PE	PERTH 07367813	Shire of Denmark, WA, Australia	MW775316-MW775319			MW820662
ides I	PERTH 07626959	Shire of Gingin, WA, Australia	MW775302-MW775306	MW820654		C00070 M TAI
	PERTH 07627025	City of Wanneroo WA, Australia	MW775307, MW775309	MW820655	MW820678	MW820664
A. compacta PE	PERTH 08700524	Shire of Wongan-Ballidu, WA, Australia	MW775275-MW775278	MW820657	MW820679	MW820665
A. griselloides PE	PERTH 09004831	City of Albany, WA, Australia	MW775270-MW775274	MW820658	$\mathbf{MW820680}$	MW820666
A. peltigera PE	PERTH 08793514	Shire of Manjimup, WA, Australia		MN909824	MN912054	
	PERTH 09138560	Shire of Mundaring, WA, Australia		MN909825	MN912055	
A. peltigera PE	PERTH 08793581	Shire of Serpentine-Jarrahdale, WA, Australia		MN909826	MN912056	MN905762
A. peltigera	AD282184	Kangaroo Island, SA, Australia		MN909828	MN912058	
A. peltigera	AD282185	Kangaroo Island, SA, Australia		MN909827	MN912057	MN905761
A. pseudoarenaria [as A. arenaria VI	VPI 364	Kalamunda, WA, Australia	GQ925388			
A. pseudoarenaria PE	PERTH 09316183	Shire of Serpentine-Jarrahdale, WA, Australia	MW795697-MW795701	MW820647	MW820673	
A. pseudoarenaria PE	PERTH 09316175	Shire of Manjimup, WA, Australia	MW795702-MW795705	MW820648		$\mathbf{MW820670}$
A. pseudoarenaria PE	PERTH 09316191	Shire of Mundaring, WA, Australia	MW795706-MW795710	MW820651	MW820676	MW820672
A. pseudoarenaria PE	PERTH 08606765	Shire of Manjimup, WA, Australia	MW775311-MW775315	MW820656	MW820681	MW820669
A. pupatju PE	PERTH 09269525	City of Melville, WA, Australia	MW775297-MW775301	MW820659		MW820668
A. pupatju PE	PERTH 09269533	City of Melville, WA, Australia	JX398329			
A. sabulosa PE	PERTH 09178694	City of Melville, WA, Australia	KF859748-KF859752			
A. sabulosa PE	PERTH 09178759	City of Melville, WA, Australia	MW775288-MW775291	MW820650	MW820675	$\mathbf{MW820660}$
A. sabulosa PE	PERTH 09178775	City of Melville, WA, Australia	MW775292-MW775296	MW820653	MW820677	MW820661
A. sabulosa	AD-C54481	Monarto Conservation Park, SA, Australia	KP137081- KP137083	MW820652		MW820667
A. wadjukiorum PE	PERTH 08793522	City of Melville, WA, Australia		MN928569	MN928570	MN928568
A. wadulawitu PE	PERTH 09144404	City of Melville, WA, Australia		MN928566	MN928560	MN928563
A. wadulawitu PE	PERTH 09144382	Shire of Esperance, WA, Australia		MN928567	MN928561	MN928564
A. wadulawitu PE	PERTH 09144390	Shire of Serpentine-Jarrahdale, WA, Australia		MN928565	MN928562	MN928562
A. djarilmari (outgroup) PE	PERTH 08776067	Shire of Cuballing, WA, Australia	KY977732	MF000750	MF000755	MF000742

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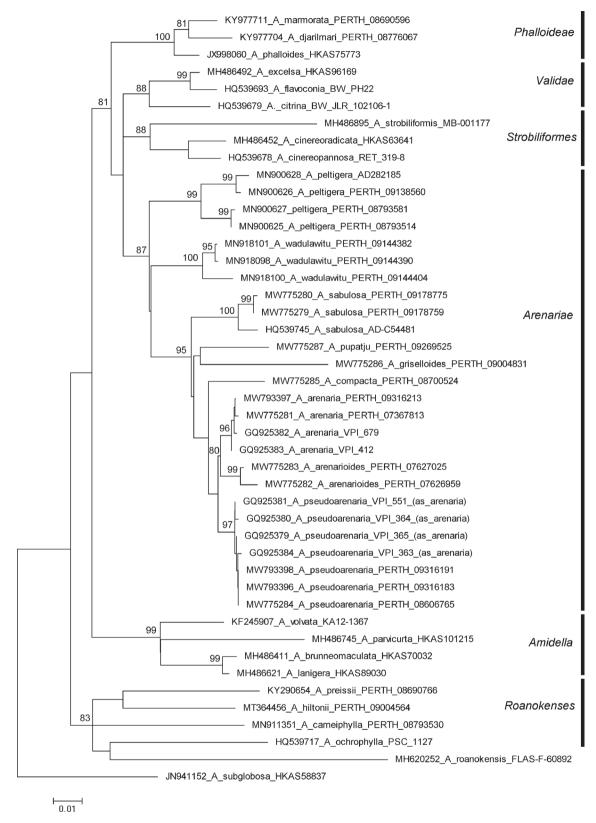


Fig. 1. Molecular phylogenetic analysis by the maximum-likelihood method of *nuLSU* (nuclear ribosomal large-subunit rRNA region) sequences (822 base pair positions), showing the position of *Amanita arenarioides*, *A. compacta*, *A. griselloides*, *A. pseudoarenaria*, *A. pupatju* and *A. sabulosa* within subgenus *Amanitina*. The tree is rooted with *A. subglobosa* (subgenus *Amanita* section *Amanita*). Maximum-likelihood bootstrap values greater than 80% are shown on the branches.

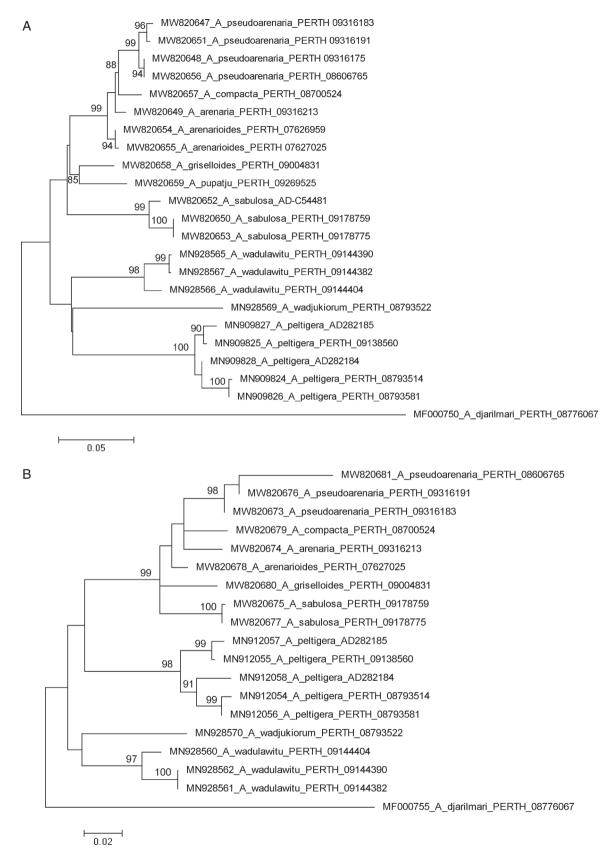
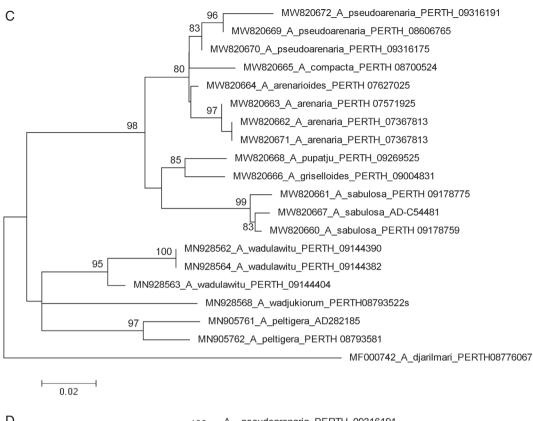


Fig. 2. Molecular phylogenetic analysis by the maximum likelihood method, of the sequences. A. $efl-\alpha$ (translation elongation-factor 1-α region) sequences (479 base pair positions). B. rpb2 RNA polymerase-II region) sequences (634 base pair positions). C. β -tubulin (β -tubulin region) sequences (362 base pair positions). D. Concatenated nuLSU, rpb2, $efl-\alpha$ and β -tubulin sequences (2241 base pair positions) of species in section Arenariae. All trees are rooted with $Amanita\ djarilmari$ (section Phalloideae). Maximum-likelihood bootstrap values greater than 80% are shown on the branches for A–C, and greater than 70% for D.



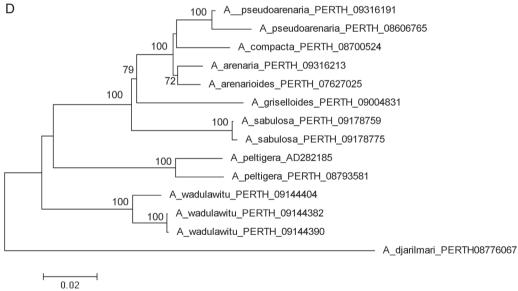


Fig. 2. (continued).

species A. arenaria, A. arenarioides, A. compacta and A. pseudoarenaria and the agaricoid species A. griselloides, A. pupatju and A. sabulosa. Taking into account the nuLSU phylogeny and the phylogenies of other regions (see below), these are considered as species-level clades and are described below (see Taxonomy section). The names of these novel taxa are indicated on the phylogenetic trees, and previous misidentifications are updated.

Additional gene regions

Additional gene regions ($ef1-\alpha$, rpb2 and β -tubulin) are available for some collections within section Arenariae (Table 2). Phylogenetic analysis of the $ef1-\alpha$ (Fig. 2A), rpb2 (Fig. 2B), β -tubulin (Fig. 2C) and concatenated nuLSU, rpb2, $ef1-\alpha$ and β -tubulin gene regions (Fig. 2D) all show that the secotioid species (A. arenaria, A. arenarioides, A. compacta and A. pseudoarenaria) together with A. griselloides, A. pupatju and

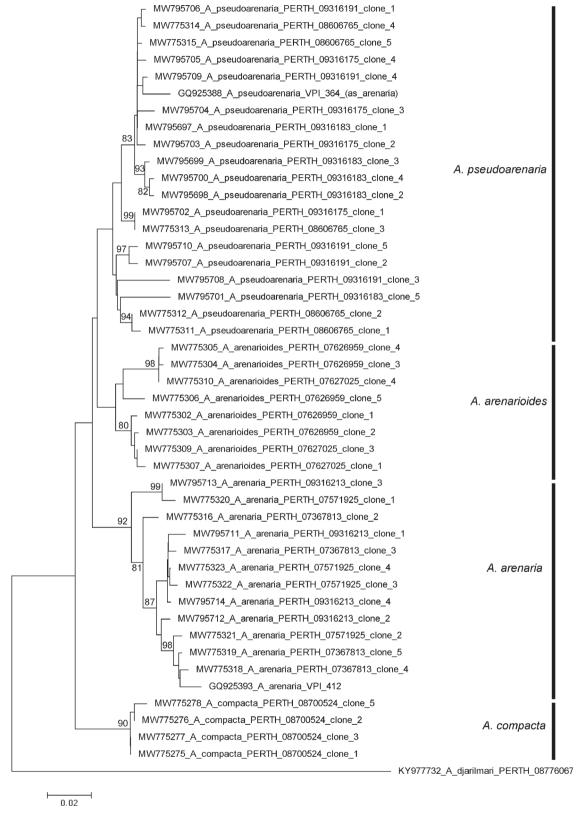


Fig. 3. Molecular phylogenetic analysis by the maximum-likelihood method of *ITS* sequences (530-base pair positions) of secotioid species from section *Arenariae*. The tree is rooted with *A. djarilmari* (section *Phalloideae*). Maximum-likelihood bootstrap values greater than 80% are shown on the branches.

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A. sabulosa form a well-supported clade, separate from A. peltigera, A. wadjukiorum and A. wadulawitu.

ITS region

Cloned haplotypes of the *ITS* region are available for both secotioid and agaricoid species from section *Arenariae*. No chimeric reads were detected in these sequences.

In the secotioid species, the *ITS* region is 569–599 base pairs long. Molecular phylogenetic analysis by the maximum-likelihood method shows that GQ925393 and GQ925388 (sequences originally identified as *A. arenaria* by Justo *et al.* 2010) are widely separated. All the new species form distinct clades, some with good bootstrap support (Fig. 3). The bootstrap support for *A. arenaria* is 92% (13 clones from four collections), for *A. arenarioides* it is 67% (8 clones from two collections), for *A. pseudoarenaria* it is 77% (20 clones from five collections) and for *A. compacta* it is 53% (4 clones from a single collection).

As can be seen from Fig. 3, there is considerable variation between the clones; they differ by 0.0–8.1% within an individual, by 0.0–9.6% among collections of the same species, and by 10.3–18.8% among different species (Table 3).

Examination of the *ITS* region shows that there is an insertion of up to 20 base pairs (AAAGGAAGCTTCACTTTTGA in MW775323) in *ITS1* in all haplotypes of *A. arenaria*; this is not present in any haplotypes of *A. pseudoarenaria*. There is a shorter, 15 base pair insertion (AGGGGAATCTTTTGA) in *ITS1* in all haplotypes of *A. arenarioides*. There is a 10 base pair deletion in *ITS1* in all haplotypes of *A. arenaria* and *A. arenarioides*, which is not present in *A. pseudoarenaria* and *A. compacta*.

In the agaricoid species *A. griselloides*, *A. pupatju* and *A. sabulosa*, the *ITS* region is 582–588 base pairs long. There is considerable variation among the clones: they differ by 0.2–9.9% among haplotypes from the same individual, by 0.7–10.1% among collections of the same species and by 12.2–18.0% among different species (Table 4).

Examination of the *ITS* region shows that there is a five base pair deletion in *ITS1* in all haplotypes of *A. griselloides* and *A. pupatju* and a four base pair insertion in *ITS1* in all haplotypes of *A. pupatju*.

Taxonomy

Section Arenariae

Cui et al. (2018, pp. 106, 117) stated that members of section Arenariae are agaricoid or sequestrate and lack clamp connections. Since their publication, more species have been recognised as being members of section Arenariae (including those detailed below), enabling us to provide a revised diagnosis and description of this section.

Diagnosis

Basiomata agaricoid or sequestrate; basiodiospores thinwalled or very slightly thickened, smooth, amyloid in agaricoid species, inamyloid in sequestrate species; clamp connections absent in all parts of the basidiome. When basidiomata is agaricoid, pileus with remnants of universal veil, lamellulae attenuate, stipe base variable. When

Table 3. Percentage difference between ITS clones from the secotioid species Amanita arenaria, A. pseudoarenaria, A. arenarioides and A. compacta

Collection	Number of clones	Amanita arenaria VPI 679 PERTH 0931621	enaria PERTH 09316213	PERTH 07367813	PERTH 07571925	Amanita pse VPI 364	Amanita pseudoarenaria VPI 364 PERTH 09316183	PERTH 09316175	PERTH 09316191	PERTH 08606765	Amanita arenarioides PERTH PERTH 07626959 07627025	narioides PERTH 07627025	Amanita compacta PERTH 08700524
A. arenaria													
VPI 679	1												
PERTH 09316213	4	3.3–6.6	0.9–5.7										
PERTH 07367813	4	1.4–6.3	2.2-6.3	0.5-5.6									
PERTH 07571925	3	1.9–7.1	0.5-6.2	8.9–6.0	0.5-6.8								
A. pseudoarenaria													
VPI 364	1	17.3	16.4 - 17.2	16.2–16.6	16.2–16.6 16.6–17.0								
PERTH 09316183	S	16.2-17.8	15.3-18.0	15.4-17.5	15.5-17.8	2.3-7.8	0.3-6.7						
PERTH 09316175	4	15.8-16.9	15.3-17.2	14.7–16.5	15.1 - 17.0	2.4-4.5	0.9-7.0	1.0-4.1					
PERTH 09316191	S	16.4 - 18.2	15.0 - 17.8	15.5-16.9	15.1 - 18.1	2.2-8.6	1.6 - 9.6	1.9-8.1	1.0 - 8.1				
PERTH 08606765	S	15.8-17.4	15.3-17.3	14.7-17.1	15.1–17.2	2.3–7.9	1.0-7.4	0.0-7.2	1.0 - 7.6	0.3–6.6			
A. arenarioides													
PERTH 07626959	5	11.6–12.7	1.6-12.7 10.5-13.0	10.2 - 12.4	10.7-13.1 15.7-16.0 14.8-16.9 14.2-15.9 15.3-18.2	15.7-16.0	14.8 - 16.9	14.2–15.9	15.3-18.2	14.2-16.8 0.3-7.2	0.3-7.2		
PERTH 07627025	3	11.9–12.9	10.7 - 13.1	10.3 - 12.1	10.8 - 13.3	15.3-16.0	14.7–16.3	13.8-15.8	14.8-18.3	13.8-16.9	0.5-8.2	0.5-6.1	
A. compacta													
PERTH 08700524	5	16.5–18.3	16.5-18.3 15.6-18.2	15.7-17.5	15.7-17.5 15.8-18.6 14.4-15.3 13.4-14.9 12.7-15.1 13.5-16.6 12.7-15.2 17.1-18.8 14.8-18.6 0.0-3.6	14.4–15.3	13.4–14.9	12.7–15.1	13.5-16.6	12.7–15.2	17.1 - 18.8	14.8–18.6	0.0–3.6

Collection	Number of clones	Amanita griselloides PERTH 09004831	Amanita pupatju PERTH 09269525	PERTH 09269533	Amanita sabulosa PERTH 09178759	PERTH 09178775	AD-C54481	PERTH 09178694
A. griselloides								
PERTH 09004831	5	0.6-5.7						
A. pupatju								
PERTH 09269525	5	12.2-13.8	0.5 - 2.0					
PERTH 09269533	1	12.8-13.8	1.4-2.7					
A. sabulosa								
PERTH 09178759	4	14.5-16.1	14.6-15.9	15.6-16.6	2.0-3.4			
PERTH 09178775	5	14.4-15.8	15.1-16.3	16.1-16.9	0.7-4.6	0.2 - 3.7		
AD-C54481	3	15.2-17.4	15.6-17.4	16.6-18.0	1.9-9.6	3.7 - 10.1	4.1 - 9.9	
PERTH 09178694	5	14.2–16.4	15.0-16.4	15.9-17.1	1.0-4.1	1.0-4.9	2.4-10.1	1.5-4.4

Table 4. Percentage difference between ITS clones from the agaricoid species Amanita griselloides, A. pupatju and A. sabulosa

basidiomata is sequestrate, gleba loculate, stipitate, basidia thin-walled, collapsing early. Known from southern Australia on sandy soil, associated with *Agonis*, *Allocasuarina*, *Corymbia*, *Eucalyptus*, *Hibbertia*, *Jacksonia*, *Melaleuca*, *Mirbelia*, *Podocarpus* and *Taxandria*.

Description

Basiomata agaricoid or sequestrate

When basidiomata agaricoid, pileus plane, margin nonstriate, appendiculate when young, context white or cream, unchanging, or becoming vinaceous-buff when bruised. Universal veil on pileus present. Lamellae adnexed to adnate, white to cream; lamellulae attenuate. Stipe subcylindric, stipe base turbinate and elongating with age or ovoid or conic. Partial veil present. Universal veil at base of stipe indistinct or as soft warts or as small, flimsy free limb. Basidiospores amyloid, thin-walled, smooth. Sub-basidial cell becoming inflated. Clamp connections absent.

When sequestrate, basidiomata hypogeus to emergent, sequestrate and stipitate; pileus subglobose or pulvinate or cylindric, white or cream, yellowing with age; margin inrolled, in contact with stipe. Universal veil on pileus usually absent. Gleba loculate, white to cream. Stipe subcylindric, apex hardly extending into gleba or extending almost to top of gleba. Partial veil absent. Universal veil at stipe base a small cup or absent. Basidiospores statismosporic, inamyloid or very slightly amyloid, wall slightly thickened, smooth. Basidia thin-walled, collapsing early, thick-walled crassobasidia sometimes present. Clamp connections absent.

Known from southern Australia on sandy soil, associated with Agonis, Allocasuarina, Corymbia, Eucalyptus, Hibbertia, Jacksonia, Melaleuca, Mirbelia, Podocarpus and Taxandria.

Amanita arenaria (O.K.Mill. & E.Horak) Justo, *Mycologia* 84(1): 65–69 (1992) (Fig. 4, 5)

Basidiomata sequestrate, secotioid, capitate, very small. Pileus white to ivory, globose to pulvinate, margin incurved. Gleba chambered, white to cream. Stipe cylindric, hardly reaching into

gleba, solid, scabrous, white. Universal veil on pileus as thin white flakes, usually missing; at stipe base a small, white to cream shallow cup. Spores ellipsoid to elongate, inamyloid. Clamp connections absent.

Type: AUSTRALIA. Western Australia: city of Albany, Two Peoples Bay Nature Reserve, 34°57′S, 118°10′E, 22 June 1989, *O.K. Miller OKM 24013*, *H.H. Miller & N.L. Bougher E697* (holo: VPI 679, iso: PERTH 07586329!).

Mycobank number: MB515037.

Published sequence

Type: *nuLSU* (GQ925382).

Illustration

Miller and Horak (1992, fig. 5–7); Bougher and Syme (1998, pp. 174–175).

Description

Basidiomata sequestrate, secotioid, capitate, very small. Pileus $5{\text -}16$ mm wide, up to 4 mm thick, white to ivory (B) without surface staining or bruising, globose to pulvinate, surface dry, silky, smooth to lumpy, margin incurved. Universal veil on pileus adnate, thin, membranous flakes, usually missing, white. Gleba white to ivory to cream (B-D), chambers round to elongate. Stipe $3{\text -}33$ mm \times $3{\text -}6$ mm, cylindric or narrowing downward, apex domed, white; surface scabrous or smooth; context solid, white to ivory (B), unchanging. Universal veil at base of stipe a small broad membranous cup up to 0.5 mm thick with smooth, dry inner surface, white to cream. Odour not distinctive or of mushrooms or coconut.

Basidiospores [160/8/8] (10–)10.5–14(–14.5) \times (6.5–)7–8.5 (–9) μ m, (L=11.4–13.4 μ m; L'=12.2 μ m; W=7.1–8.2 μ m; W'=7.5 μ m; Q=(1.25–)1.44–1.86(–2.00); Q=1.54–1.70; Q'=1.62), colourless, thick-walled, smooth, inamyloid or very slightly amyloid, ellipsoid to elongate, contents monoguttulate; apiculus terminal occasionally sublateral, tapered or cylindric, 0.5–1 \times 1–1.5 μ m, rounded.

Peridium up to 20 μm thick, colourless to pale yellow, not highly gelatinised; filamentous hyphae 3–15 μm wide, walls



Fig. 4. Amanita arenaria (PERTH 07367813).

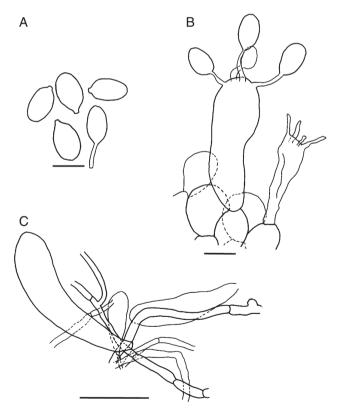


Fig. 5. Microscopic features of *Amanita arenaria*. A. Basidiospores. B. Basidia and sub-basidial cells. C. Universal veil from pileus, scalp section gently squashed. Scale bars: 10 μm (A, B) 50 μm (C). (A, B: PERTH 07586329, isotype; C: PERTH 07367813).

thickened, colourless, periclinal orientation, gelatinising; inflated cells and vascular hyphae not seen. Pileus context consisting of dominant or equal filamentous hyphae, inflated cells and very infrequent vascular hyphae; filamentous hyphae 3-15 µm wide, with widest hyphae constricted at septa, slightly thickened walls, colourless, gelatinising; inflated cells up to $120 \times 25 \mu m$, walls thin or slightly thickened, gelatinising, clavate or ovoid or ventricose or ellipsoidal or spherical or cylindrical, colourless; vascular hyphae 2–7 µm wide, occasionally branched, pale yellow. Dissepiments of dominant filamentous hyphae and inflated cell (vascular hyphae not seen); filamentous hyphae 4-30 µm wide, with widest hyphae constricted at septa, walls thin or slightly thickened, colourless, gelatinising; inflated cells up to $110 \times 35 \mu m$, walls thin or slightly thickened, gelatinising, clavate or ovoid or ventricose or ellipsoidal or spherical, colourless. Subhymenium cellular, up to 40 µm wide, of up to 4 cell layers, with basidia arising terminally from ovoid or clavate or globose terminal segments up to 13 µm wide. Basidia: [30/2/2] (23–)24–46(–50) × 9–14 µm, thin-walled, colourless, collapsing early, ~67% 4-spored, ~13% 3-spored, ~20% 2-spored, sterigmata up to 15 \times 2 μ m, cylindrical; crassobasidia present. Universal veil on pileus not layered, elements with periclinal orientation, consisting of filamentous hyphae and frequent to very infrequent inflated cells (vascular hyphae not observed); filamentous hyphae 4–12 µm wide, with thin or slightly thickened walls, colourless, gelatinising; inflated cells up to $80 \times 20 \,\mu m$ when ventricose or up to $150 \times 35 \,\mu m$ when clavate or up to $20 \times 10 \mu m$ when ovoid, with thin or slightly thickened walls, terminal, colourless, gelatinising. Universal veil at stipe base layered, elements with axial

orientation. Outer layer narrow, compact, consisting of filamentous hyphae (inflated cells and vascular hyphae not observed), 3-10 µm wide, thick-walled, colourless or pale yellow, gelatinising, often interlocking. Inner layer wide, loose, consisting of dominant or equal filamentous hyphae, inflated cells and very infrequent vascular hyphae; filamentous hyphae 4-17 µm wide, thick-walled, colourless or pale yellow, gelatinising; inflated cells up to $85 \times 25 \,\mu m$ when clavate or up to $85 \times 50 \,\mu\text{m}$ when ovoid or up to $50 \times 50 \,\mu\text{m}$ when spherical, thin-walled, colourless, gelatinising; vascular hyphae 4 µm wide, pale yellow. Stipe context longitudinally acrophysalidic, consisting of filamentous hyphae, dominant acrophysalides and very infrequent vascular hyphae; filamentous hyphae 2–10 µm wide, colourless or pale yellow, gelatinising; acrophysalides up to $400 \times 40 \,\mu m$ clavate, terminal, colourless or pale yellow, gelatinising; vascular hyphae 2–15 µm wide, occasionally branched, pale yellow. Clamp connections not seen.

Habit, habitat and distribution

Gregarious or scattered, buried in sandy soil, with only the pileus showing. In native vegetation, nearby plants include *Agonis flexuosa*, *A. parviceps*, *Allocasuarina fraseriana*, *Corymbia calophylla*, *Eucalyptus marginata*, *E. megacarpa*, *Hakea* sp., *Jacksonia horrida*, *Podocarpus drouynarius* and *Taxandria parviceps*. Occurs in the Warren WAR01 IBRA region (Department of the Environment 2013). Fruiting period is June to August.

Notes

On the basis of their nuLSU sequences and spore shape, Justo $et\ al.\ (2010)$ suggested that the $A.\ arenaria$ collections of Miller and Horak (1992) represent two cryptic species. Our analysis supports this conclusion. The nuLSU sequence

GQ925382 from the holotype of *A. arenaria* forms a well-supported clade with other collections from the City of Albany and shire of Denmark, on the southern coast of WA (Fig. 1); their β -tubulin and ITS sequences also form well supported clades when compared with other secotioid species (Fig. 2C, 3). All collections of *A. arenaria* have ellipsoid to elongate spores ($\mathbf{Q} = 1.54-1.70$).

Specimens examined

WESTERN AUSTRALIA. City of Albany, O.K. 24012, H.H. Miller & N.L. Bougher E696 (PERTH 07586310); loc. id., O.K. Miller 24018 & H.H. Miller (PERTH 02241757); loc. id., O.K. 24012, H.H. Miller & N.L. Bougher E696 (PERTH 07586310); loc.id., K. Syme 87/91 & M. Hart (PERTH 04987322); loc. id., K. Syme 428/92 & M. Hart (PERTH 05372178); loc.id., K. Syme 753/94, M. Hart & N.L. Bougher H 7333 (PERTH 05505968); loc.id., KS 3059 (PERTH 09316213); shire of Denmark, K. Syme 1361/04 (PERTH 07367813, MEL 2292314); loc. id., I.C. Tommerup, K. Syme, M.E. Bougher, N.L. Bougher H 7362 (PERTH 07571925).

Amanita pseudoarenaria E.M.Davison, Giustiniano & Bougher, sp. nov.

(Fig. 6, 7)

Basidiomata sequestrate, secotioid, capitate, very small. Pileus white to cream, sometimes yellow stained, globose to pulvinate, umbilicate. Gleba chambered, white to cream. Stipe cylindric, hardly reaching into gleba, solid, scabrous, white to cream, sometimes staining yellow. Universal veil a shallow cup at stipe base, white to cream. Spores subglobose to broadly ellipsoid, inamyloid. Clamp connections absent.

Type: AUSTRALIA. Western Australia: shire of Manjimup, 34°10′30″S, 116°35′41″E, 2 June 2013, *N.L. Bougher 1072* (holo: PERTH 08606765!).

Mycobank number: MB839131.



Fig. 6. Amanita pseudoarenaria (PERTH 08606765, holotype).

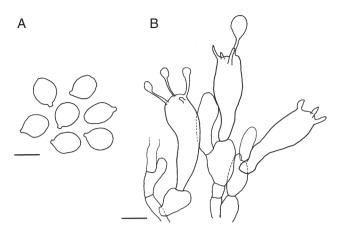


Fig. 7. Microscopic features of *Amanita pseudoarenaria*. A. Basidiospores. B. Basidia and sub-basidial cells. Scale bars: $10~\mu m$. (PERTH 08606765, holotype).

Published sequences

Type: *ITS* (MW775312), *nuLSU* (MW775284), *ef1*-α (MW820656), *rpb2* (MW820678), *β-tubulin* (MW820669).

Description

Basidiomata sequestrate, secotioid, capitate, very small. Pileus 5-15 mm wide, up to 5 mm thick, white to ivory to cream (B–D; 4A1-4A2), sometimes yellow stained, globose to pulvinate, umbilicate, surface smooth to lumpy. Universal veil on pileus absent. Gleba white to ivory to cream (B–D; 4A1-4A2), chambers elongate. Stipe $5-33 \times 3-8$ mm, cylindric or narrowing upward or narrowing downward, apex domed, white to ivory (B), sometimes yellow stained, surface scabrous or smooth; context solid, white. Universal veil at base of stipe a cup up to 5 mm high and up to 1 mm thick, white to ivory (B) often missing. Odour none.

Basidiospores [160/8/7] (8.5–)9–12(–14) \times (7–)7.5–10 (–11) μ m, (L = 9.9–11.0 μ m; L' = 10.6 μ m; W = 8.1–9.2 μ m; W' = 8.7 μ m; Q = (1.05–)1.06–1.37(–1.60); Q = 1.11–1.27; Q' = 1.22), colourless, thick-walled, smooth, inamyloid or very slightly amyloid, subglobose to broadly ellipsoid, contents monoguttulate; apiculus terminal, occasionally sublateral, tapered or cylindric, 1–2 \times 1–2 μ m, rounded.

Peridium up to 50 µm thick, colourless; filamentous hyphae 2-15 µm wide, walls slightly thickened, hyaline, contents colourless or very pale yellow, periclinal orientation, gelatinising; inflated cells not seen; vascular hyphae very infrequent, 6 µm wide, pale yellow. Pileus context consisting of dominant, equal or less frequent filamentous hyphae, inflated cells and very infrequent vascular hyphae; filamentous hyphae 2-15 µm wide, with widest hyphae constricted at septa, slightly thickened walled, colourless, gelatinising; inflated cells up to $135 \times 30 \mu m$ thin-walled, clavate or ovoid or ventricose or ellipsoidal or spherical, colourless, gelatinising; vascular hyphae 5–8 µm wide, pale yellow. Dissepiments of filamentous hyphae 3-15 μm wide, walls thin or slightly thickened, colourless; inflated cells and vascular hyphae not seen. Subhymenium ramose, with basidia arising terminally from barely inflated to barrel-shaped terminal segments up to 12 µm wide. Basidia [55/4/4] (18–) $30-47(-54) \times (6-)9-13(-15)$ µm, thin-walled, colourless, collapsing early, ~42% 4-spored, ~29% 3-spored, ~22% 2-spored, \sim 7% 1-spored, sterigmata up to 10 \times 2 μ m, cylindrical; crassobasidia present. Universal veil at stipe base layered, elements with axial orientation. Outer layer narrow, elements sometimes interlocking, consisting of dominant filamentous hyphae and very infrequent inflated cells (vascular hyphae not seen); filamentous hyphae 2-10 µm wide, thick-walled, colourless or pale yellow, gelatinising; inflated cells up to 90 × 25 µm when clavate or up to 35 \times 18 μm when ovoid or up to 30 \times 30 μm when spherical, colourless. Inner layer wide, loose, consisting of dominant or frequent, inflated cells and very infrequent vascular hyphae; filamentous hyphae 3-15 µm wide, with widest hyphae constricted at septa, colourless or pale yellow, gelatinising; inflated cells up to 300 \times 35 μ m when clavate or 75 \times 55 μm when ovoid or 40 \times 40 μm when spherical or 35 \times 18 μm when ellipsoidal or 140 \times 50 μm when ventricose or 40 \times 20 μm when pyriform, colourless or pale yellow, gelatinising; vascular hyphae 4-12 µm wide, pale yellow. Stipe context longitudinally acrophysalidic, consisting of filamentous hyphae, dominant acrophysalides and very infrequent vascular hyphae; filamentous hyphae 3–12 µm wide, colourless, gelatinising; acrophysalides up to 440 \times 35 μm when clavate or 375 \times 35 μ m when cylindric or 150 \times 35 μ m when ventricose, colourless or pale yellow, gelatinising; vascular hyphae 2-7 µm wide, occasionally branched, pale yellow. Clamp connections not seen.

Habit, habitat and distribution

Gregarious, initially buried in sand or clayey sand or clayey lateritic gravel or loam, emerging as the stipe elongates. In native vegetation, nearby plants include *Agonis linearifolia*, *A. parviceps, Corymbia calophylla, Eucalyptus marginata*, *E. patens, Hibbertia* sp. and *Mirbelia dilatata*. Occurs in the Swan Coastal Plain Perth SWA02, Northern Jarrah Forest JAF01, Southern Jarrah Forest JAF02, and Warren WAR01 IBRA subregions (Department of the Environment 2013). Fruiting period is May to August.

Etymology

From the Greek *pseudo-* meaning 'false', referring to its similarity to *A. arenaria*.

Notes

This is the cryptic secotioid species, which was discussed by Justo *et al.* (2010). The *nuLSU* sequences form a well supported clade, as do *ef1-\alpha*, *rpb2*, β -tubulin and *ITS* sequences that differ from those of *A. arenaria* (Fig. 1, 2A–C, 3, Table 3). This species is widespread in the southwest of WA. It is, in macroscopic appearance, similar to *A. arenaria*, but can be distinguished from it microscopically by its subglobose to broadly ellipsoid ($\mathbf{Q} = 1.11-1.27$), not ellipsoid to elongate ($\mathbf{Q} = 1.54-1.70$) spores. It differs from secotioid species in section *Amarrendiae*, and from *A. torrendii* Justo because of the absence of clamp

connections. On this basis A. pseudoarenaria is described as a new species.

Some collections produce a bright yellow liquid that dries as a yellow stain where they have been damaged by insects (Fig. 6). This is not specific to *A. pseudoarenaria* because it has been observed on some collections of at least one additional, yet to be described, secotioid species from this section (E. M. Davison, pers. obs.).

Specimens examined

WESTERN AUSTRALIA. City of Albany, K. Syme 428/92 (PERTH 05372178); shire of Busselton, E.M. Davison 61-2015 & P.J.N. Davison (PERTH 09316167); shire of Manjimup, E.M. Davison 20-2013 & P.J. N. Davison (PERTH 09316175); shire of Mundaring, E.M. Davison 46-2015 & P.J.N. Davison (PERTH 09316191); shire of Serpentine–Jarrahdale, E.M. Davison 19-2014 & P.J.N. Davison (PERTH 09316183); loc. id. E.M. Davison 25-2020 & P.J.N. Davison (PERTH 09316205).

Amanita arenarioides Bougher, E.M.Davison & Giustiniano, sp. nov.

(Fig. 8, 9)

Basidiomata sequestrate, secotioid, capitate, very small. Pileus white or ivory white, bruising dull yellow, globose to pulvinate, margin incurved. Gleba chambered, white or ivory. Stipe cylindric, hardly reaching into gleba, solid, white, staining yellow. Universal veil on pileus not seen; at stipe base a small, shallow cup. Spores elongate to cylindric, inamyloid. Clamp connections absent.

Type: AUSTRALIA. City of Wanneroo, Yanchep National Park, 31°30′S, 115°42′E, 13 June 1997, *N.L. Bougher & R. T. Wills H 7501* (holo: PERTH 07627025!).

Mycobank number: MB839132.

Published sequences

Type: *ITS*: (MW775309), nuLSU (MW775283), $ef1-\alpha$ (MW820655), rpb2 (MW820678), β -tubulin (MW820664).

Description

Basidiomata sequestrate, secotioid, capitate, very small. Pileus $5{\text -}16$ mm wide, up to 8 mm thick, white, or ivory white, bruising dull yellow, globose to pulvinate, margin incurved, clasping, surface dry. Gleba chambered, chambers rounded, white or ivory. Stipe $7{\text -}15 \times 2{\text -}6$ mm, cylindric, white bruising dull yellow, surface smooth or with recurved scales; context solid, white. Universal veil a small basal cup up to 3 mm high. Odour none

Basidiospores [160/8/2] (9–)10.5–14(–15) \times (5–)5.5–7 µm, (L=10.7–12.8 µm, L'=12.2 µm; W=5.8–6.5 µm, W'=6.2 µm; Q = (1.50–)1.69–2.33(–2.60); Q = 1.85–2.13, Q' = 1.99) colourless, wall slightly thickened, smooth, not adaxially flattened, inamyloid or very slightly amyloid, elongate to cylindric, contents monoguttulate; apiculus terminal or slightly sublateral, tapered, \sim 1 \times 1.5 µm, rounded or truncate.

Peridium up to 10 μ m thick, not continuous, very pale yellow; filamentous hyphae 3–5 μ m wide, walls slightly thickened, periclinal orientation, gelatinising; inflated cells not observed; vascular hyphae 6 μ m wide, pale yellow. Pileus context consisting of dominant or frequent filamentous hyphae, inflated cells and very infrequent vascular hyphae; filamentous hyphae 4–18 μ m wide, with widest hyphae constricted at septa, thin or slightly thickened walls, colourless, gelatinising; inflated cells up to $70 \times 30 \ \mu$ m, thin or slightly thickened walls, ellipsoid or clavate or ovoid or cylindric, terminal or in chains of 2; vascular hyphae 3–5 μ m wide, pale yellow. Dissepiments of dominant filamentous hyphae encircling chambers, frequent to infrequent inflated

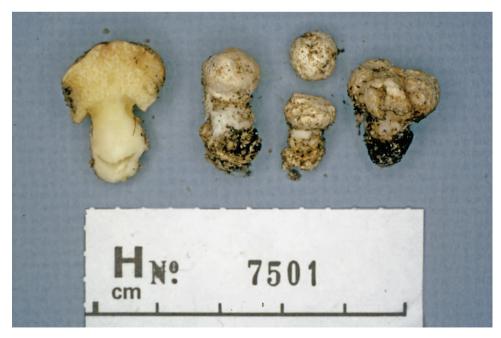
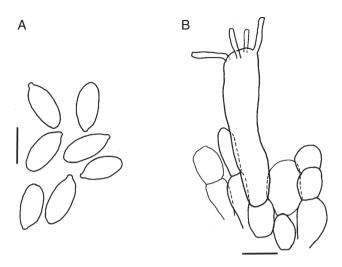


Fig. 8. Amanita arenarioides (PERTH 07627025, holotype).



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Fig. 9. Microscopic features of *Amanita arenarioides*. A. Basidiospores. B. Basidium and sub-basidial cells. Scale bars: $10~\mu m$. (PERTH 07627025, holotype).

cells and very infrequent vascular hyphae; filamentous hyphae 3–12 µm wide, thin-walled, colourless, gelatinising; inflated cells up to $50 \times 20 \mu m$ thin or slightly thickened walls, ovoid or ventricose or clavate or cylindrical or spherical, colourless; vascular hyphae 4-6 µm wide, pale yellow. Subhymenium inflated, with basidia arising terminally from segments up to 15 μ m wide, basidioles frequent. Basidia [24/2/2] (30–)32–45 \times (8-)9-12 µm, thin-walled, collapsing, colourless $\sim 67\%$ 4-spored, \sim 17% 3-spored, \sim 8% 2-spored, \sim 8% 1-spored, sterigmata up to $10 \times 2 \mu m$. Universal veil on pileus absent. Universal veil at stipe base layered with axial orientation; outer layer very narrow, of filamentous hyphae (inflated cells and vascular hyphae not observed), 3–15 μm wide, thick, pale yellow walls, gelatinising; inner layer wide, of dominant filamentous hyphae and frequent inflated cells (vascular hyphae not observed), filamentous hyphae 2-15 µm wide, thick, pale yellow gelatinising walls; inflated cells up to $85 \times 25 \mu m$ when clavate or $70 \times 60 \ \mu m$ when ellipsoidal or $40 \times 35 \ \mu m$ when ovoid or to 75 \times 20 μm when ventricose or 25 \times 25 μm when spherical, thick, pale vellow gelatinising walls. Stipe context longitudinally acrophysalidic, consisting of frequent filamentous hyphae, dominant acrophysalides and very infrequent vascular hyphae; filamentous hyphae 3-12 µm wide, slightly thickened, pale yellow gelatinising walls; acrophysalides up to 240 × 40 µm, slightly thickened, pale yellow gelatinising walls, clavate; vascular hyphae 3–8 μm wide, occasionally branched, pale yellow. Clamp connections not seen.

Habit, habitat and distribution

Gregarious, barely emerging from sand. In native vegetation, nearby plants include *Banksia grandis*, *Eucalyptus marginata*. Occurs in the Swan Coastal Plain SWA02 IBRA subregion (Department of the Environment 2013). Fruiting period is June to July.

Etymology

From the Latin *-oideus* meaning 'like', referring to its similarity to *A. arenaria*.

Notes

The nuLSU sequences form a well-supported clade, as do $ef1-\alpha$ and ITS sequences that differ from those of the secotioid species A. arenaria and A. pseudoarenaria (Fig. 1, 2A–C, 3, Table 3). This species has been found only north of Perth. It is, in macroscopic appearance, similar to A. arenaria and A. pseudoarenaria, apart from the basidiomes slowly developing a dull yellowish colour after handling. It can be distinguished from A. arenaria and A. pseudoarenaria microscopically by its elongate to cylindric ($\mathbf{Q} = 1.85-2.13$), not ellipsoid to elongate ($\mathbf{Q} = 1.54-1.70$, A. arenaria) or subglobose to broadly ellipsoid ($\mathbf{Q} = 1.11-1.27$, A. pseudoarenaria), spores. It differs from secotioid species in section Amarrendiae, and from A. torrendii because of the absence of clamp connections. On this basis, A. arenarioides is described as a new species.

Specimens examined

WESTERN AUSTRALIA. Shire of Gingin, *I.C.P. Tommerup H* 7508 (PERTH 07626959).

Amanita compacta Bougher, E.M.Davison & Giustiniano, sp. nov.

(Fig. 10, 11)

Basidiomata sequestrate, secotioid, cylindric, very small. Pileus white, unchanging, thimble-shaped, thin, clasping. Gleba chambered, white or ivory. Stipe cylindric, almost reaching top of pileus, solid, white. Universal veil absent. Spores inamyloid, broadly ellipsoid to ellipsoid. Clamp connections absent.

Type: AUSTRALIA. Western Australia: Shire of Wongan–Ballidu, Elphin Nature Reserve, 30°51′43″S, 116°41′34″E, 10 Aug. 2015, *N.L. Bougher 1180* (holo: PERTH 08700524!).

Mycobank number: MB839133.

Published sequences

Type: *ITS* (MW775277), *nuLSU* (MW775285), *ef1-* α (MW820657), *rpb2* (MW820679), β -tubulin (MW820665).

Description

Basidiomata sequestrate, secotioid, cylindric, very small. Pileus 7–11 mm wide, up to 3 mm thick, white, without surface staining or bruising, thimble-shaped, covering stipe apex, clasping, surface dry. Gleba with elongate chambers, white or ivory. Stipe $15-18 \times 5-11$ mm, cylindric tapered within pileus, white, surface smooth; context solid, white. Partial veil absent. Universal veil absent on pileus and at stipe base. Odour none.

Basidiospores [80/4/1] 9–12(–13) \times (6–7.5(–8.5) μ m, (L = 1.40–1.63 μ m; L' = 10.6 μ m; W = 6.8–7.2 μ m; W' = 7.0 μ m; Q = (1.18–)1.33–1.83(–1.92); Q = 1.40–1.63; Q' = 1.53) colourless, slightly thickened wall, smooth, not adaxially flattened, some distorted, inamyloid or very slightly amyloid, broadly ellipsoid to ellipsoid to elongate, contents monoguttulate; apiculus terminal or slightly sublateral, cylindric or tapered, \sim 1 \times 1.5 μ m, truncate.



Fig. 10. Amanita compacta (PERTH 08700524, holotype).

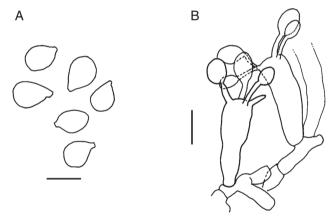


Fig. 11. Microscopic features of *Amanita compacta*. A. Basidiospores. B. Basidia and sub-basidial cells. Scale bars: $10 \mu m$. (PERTH 08700524, holotype).

Peridium present, not continuous, colourless; filamentous hyphae 4–8 μm wide, with some terminal segments up to 10 μm , walls slightly thickened, periclinal orientation, gelatinising; inflated cells not observed. Pileus context consisting of dominant filamentous and infrequent or very infrequent inflated cells; filamentous hyphae 3–12 μm wide, walls slightly thickened, interlocking in places, colourless, gelatinising; inflated cells up to 70 \times 15 μm , slightly thickened walls, clavate or ovoid or ventricose, colourless, gelatinising. Dissepiments of dominant filamentous hyphae encircling chambers and very infrequent inflated cells; filamentous hyphae 3–7 μm wide, thin-walled, colourless;

inflated cells up to 55 \times 12 μm clavate, colourless. Subhymenium ramose, with basidia arising terminally from segments 4 μm wide, becoming inflated, up to 7 μm . Basidia [20/1/1] (20–)25–36(–37) \times (7–)8–11(–15) μm , thin-walled, collapsing, colourless $\sim\!65\%$ 4-spored, $\sim\!20\%$ 3-spored, $\sim\!15\%$ 2-spored, sterigmata up to 7 \times 2 μm . Stipe context longitudinally acrophysalidic, consisting of frequent filamentous hyphae and dominant acrophysalides; filamentous hyphae 2–6 μm wide, slightly thickened walls, colourless, gelatinising; acrophysalides up to 300 \times 35 μm , slightly thickened walls, clavate or cylindric, colourless, gelatinising. Vascular hyphae not seen. Clamp connections not seen.

Habit, habitat and distribution

Gregarious, barely emerging from sand. In native vegetation, nearby plants *Allocasuarina campestris*. Occurs in the Lesueur Sandplain GES02 IBRA subregion (Department of the Environment 2013). Fruiting period is August.

Etymology

From the Latin *compactus* meaning 'close-packed', referring to the compact, rather stumpy appearance of the basidiomata.

Notes

Only one collection of this distinctive species is available. It differs macroscopically from the secotioid species *A. arenaria*, *A. pseudoarenaria* and *A. arenarioides* because the basidiomes are cylindric, not capitate, and the gleba is thimble-shaped, not globose to pulvinate. Also, the universal veil is absent. Microscopically it differs from *A. pseudoarenaria* and

A. arenarioides because the spores are ellipsoid to elongate ($\mathbf{Q}=1.40$ –1.63), not subglobose to broadly ellipsoid ($\mathbf{Q}=1.11$ –1.27, A. pseudoarenaria) or elongate to cylindric ($\mathbf{Q}=1.85$ –2.13). It differs from secotioid species in section Amarrendiae, and from A. torrendii because of the absence of clamp connections. The nuLSU, ef1- α and ITS sequences differ from those of A. arenaria, A. pseudoarenaria and A. arenarioides (Fig. 1, 2A 3, Table 3). On this basis, A. compacta is described as a new species.

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Amanita pupatju E.M.Davison, Giustiniano, McGurk & E.L.J.Watkin, sp. nov.

(Fig. 12, 13)

Basidiomata agaricoid, very small to large. Pileus white. Universal veil on pileus white, felted to floccose patches over disc, not evident at base of stipe. Lamellae white. Stipe white with mealy or floccose surface and conic bulb. Partial veil arachnoid, superior, white. Spores amyloid,





Fig. 12. Amanita pupatju (PERTH 09269525, holotype).

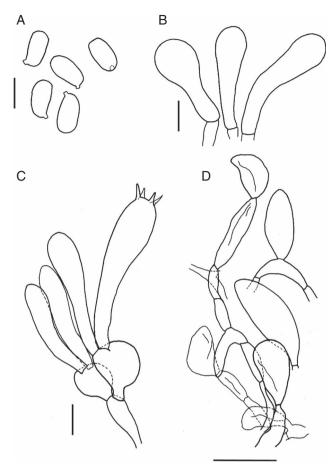


Fig. 13. Microscopic features of *Amanita pupatju*. A. Basidiospores. B. Marginal cells. C. Basidia and sub-basidial cells. D. Universal veil on pileus. Scale bars: 10 μm (A, B, C) 50 μm (D). (PERTH 09269525, holotype).

elongate to cylindric. Universal veil not layered, with no dominant orientation, composed of terminal inflated cells that are occasionally in short chains. Clamp connections absent.

Type: AUSTRALIA. Western Australia: city of Melville, Murdoch University grounds, 32°47′27″S, 115°50′04″E, 31 May 2015, *E.M. Davison 9–2015 & P.J.N. Davison*, (holo: PERTH 09269525!).

Mycobank number: MB839134.

Published sequences

Type: *ITS* (MW775301), *nuLSU* (MW775287), *ef1-* α (MW820659), β -*tubulin* (MW820668).

Description

Pileus 22–115 mm wide, up to 6 mm thick, white to very pale clay pink (6A2) with age, without surface staining or bruising, initially convex becoming plane with slightly depressed centre, surface tacky when moist; margin non-striate, appendiculate. Universal veil on pileus adnate, thin, felted to floccose, breaking into small patches over disc, white. Lamellae adnexed to adnate, subcrowded, white, 5–10 mm

broad, margin concolorous, fimbriate; lamellulae plentiful to infrequent in several lengths, rounded truncate or truncate or rounded attenuate or attenuate. Stipe 35–75 mm long, 8–40 mm wide, cylindrical or tapering upward or tapering downward, white, surface mealy or floccose or fibrillose. Partial veil superior, arachnoid, white. Bulb 14–35 × 16–39 mm, conic to tapered, occasionally ovoid when young, white to grey. Remains of universal veil at top of bulb not evident, or as a slight ridge. Pileus and stipe context white, unchanging, stipe solid becoming hollow. Smell none when young, ammoniac when old. Spore deposit white or B (ivory).

Basidiospores [200/10/8] (8.5–)9.5–13.5(–17) \times (4.5–)5–7 (–7.5) μ m, (L = 10.6–13.6 μ m; L' = 11.5 μ m; W = 5.2–6.5 μ m; W' = 5.8 μ m; Q = (1.55–)1.67–2.33(–3.00); Q = 1.80–2.12; Q' = 1.98), colourless, thin-walled, smooth, amyloid, ellipsoid to elongate to cylindrical, contents monoguttulate or granular; apiculus sublateral, cylindric or tapered, ~1 \times 1 μ m, rounded or truncate.

Pileipellis up to 300 µm thick in old specimens, with a colourless gelatinised suprapellis up to 200 µm thick and colourless subpellis, consisting of filamentous hyphae and infrequent vascular hyphae (inflated cells not observed). filamentous hyphae 3-15 µm wide, thick-walled, walls hyaline, gelatinising, contents colourless, radially orientated with some interweaving; vascular hyphae 3-6 µm wide, pale yellow or yellowish-brown. Pileus context consisting of filamentous hyphae (dominant), inflated cells and very infrequent vascular hyphae, filamentous hyphae 3-40 µm wide, with widest hyphae constricted at septa, thin-walled, colourless; inflated cells to $250 \times 40 \mu m$, thin-walled, clavate or ventricose or ovoid, terminal, colourless; vascular hyphae 3–11 µm wide, pale yellow. Lamella trama bilateral, divergent. Central stratum up to 40 µm wide, consisting of filamentous hyphae (inflated cells and vascular hyphae not observed), filamentous hyphae 3-10 µm wide, thin-walled, colourless. Subhymenial base with angle of divergence ~30° from central stratum, with filamentous hyphae following smooth broad curve to subhymenium, consisting of dominant filamentous hyphae, inflated cells (infrequent to frequent) and very infrequent vascular hyphae; filamentous hyphae 3-20 µm wide, widest close to subhymenium and constricted at septa, frequently branched, thin-walled, colourless; inflated cells up to 200 \times $20 \,\mu\text{m}$ when clavate or up to $160 \times 12 \,\mu\text{m}$ when ventricose or up to $50 \times 18 \,\mu m$ when elliptic or up to $60 \times 20 \,\mu m$ when cylindric, terminal, colourless; vascular hyphae 3–5 µm wide, occasionally branched, pale greenish-yellow. Subhymenium with basidia arising terminally from pyriform to spherical terminal segments up to 24 µm wide. Lamella edge tissue sterile, with inflated cells infrequent to frequent, up to 40×11 µm when clavate or up to 35 \times 11 μ m when capitate or up to 30 \times 14 μ m when broadly clavate or up to $30 \times 14 \,\mu\text{m}$ when pyriform or up to $27 \times 7 \mu m$ when cylindric, colourless. Basidia [140/7/7] (35–) $38-57(-65) \times (9-)10-13(-14)$ µm, thin-walled, colourless, \sim 93% 4-spored, \sim 3% 3-spored, \sim 4% 2-spored, sterigmata to $8 \times 2 \,\mu\text{m}$. Universal veil on pileus not layered, elements with no dominant orientation, consisting of frequent to dominant filamentous hyphae, dominant to frequent inflated cells and very infrequent vascular hyphae, filamentous hyphae 5–15 μm wide, with widest hyphae sometimes constricted at septa, colourless, some gelatinising; inflated cells up to $90\times75~\mu m$ when ovoid or up to $220\times55~\mu m$ when ventricose or up to $100\times100~\mu m$ when spherical or up to $80\times60~\mu m$ when elliptic or up to $95\times25~\mu m$ when cylindrical, terminal or in chains of up to 3, colourless, some gelatinising; vascular hyphae 6–12 μm wide, occasionally branched, pale yellow. Universal veil at stipe base absent. Stipe context longitudinally acrophysalidic, consisting of frequent filamentous hyphae, dominant acrophysalides and very infrequent to frequent vascular hyphae, filamentous hyphae 2–8 μm wide, colourless; acrophysalides to $420\times35~\mu m$ clavate or cylindric, terminal, colourless or pale yellow, gelatinising; vascular hyphae 3–10 μm wide, occasionally branched, pale yellow or pale brownish-yellow. Partial veil not examined. Clamp connections not observed.

Habit, habitat and distribution

Singly or gregarious in sand. In degraded native vegetation, nearby plants include *Corymbia calophylla* and *Eucalyptus marginata*. Occurs in the Swan Coastal Plain Perth SWA02 IBRA subregion (Department of the Environment 2013). Fruiting period is April to June.

Etymology

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Pupatju means 'to cover the head' in the Nhanda dialect of the Western Australian Aboriginal Noongar language, reflecting the patches of universal veil on the pileus. The epithet is formed as a noun in apposition.

Notes

Figure 1 shows A. pupatju is a member of section Arenariae. There are several species with a white or pale pileus, in which clamp connections are either absent or very infrequent. Amanita pupatju has similar shaped, but longer, spores than does A. preissii (A. pupatju [200/10/8] $L = 10.6-13.6 \mu m$, $W = 5.2-6.5 \mu m$, Q = 1.80-2.12; A. preissii [340/11/11] $L = 8.8-11.5 \mu m$, $W = 5.2-6.0 \mu m$, Q = 1.65-2.13), but the pileus, universal veil and partial veil do not age pale cinnamon to saffron (Davison et al. 2017b). It has spores of shape and size similar to those of A. lesueurii ([200/10/5] L = 11.1–12.4 μ m, $W = 5.5-6.2 \mu m$, Q = 1.90-2.26), but lacks the felted or floccose universal veil on the pileus which becomes vinaceous-buff to grey with age (Davison et al. 2013). Amanita pupatju has spores that are wider than those of A. wadulawitu ([859/21/ 21] L = 10.2–12.5 μ m, W = 4.4–5.2 μ m, Q = 2.21–2.55) and a much smaller and less well defined basal bulb (McGurk et al. 2016). It differs from A. clelandii (E.-J. Gilbert) E.-J. Gilbert, and A. variabilis (E.-J. Gilbert) E.-J. Gilbert & Cleland, two poorly known species from South Australia, by its larger size and different-shaped spores (Reid 1980). The nuLSU sequence, efl- α , β -tubulin show that it clusters with A. griselloides (Fig. 1, 2A, 2C), but the ITS sequences differ by 12.2–13.8% (Table 4). Amanita pupatju differs from A. griselloides because it has a white, not pale grey or brown, pileus and the inflated cells in the universal veil on the pileus have colourless, not brown, contents (Reid 1980; see below). On this basis, Amanita pupatju is described as a new species.

Specimens examined

WESTERN AUSTRALIA. City of Melville, E.M. Davison 22-1990 & P.J.N. Davison (PERTH 03097129); loc. id., E.M. Davison 37-1990 & P.J.N. Davison (PERTH 03096955); loc. id., E.M. Davison 52-1990 & P.J.N. Davison (PERTH 03097080); loc. id., E.M. Davison 8-1991 & P.J.N. Davison (PERTH 03096726); loc. id., E.M. Davison 11-1991 & P.J.N. Davison (PERTH 03096947); loc. id., E.M. Davison 15-1992 & P.J.N. Davison (PERTH 03096998); loc. id., E.M. Davison 3-1995 & P.J.N. Davison (PERTH 09269479); loc. id., L.E. McGurk 41–2005 (PERTH 09269533).

Amanita sabulosa E.M.Davison & Giustiniano, sp. nov.

(Fig. 14, 15)

Basidiomata agaricoid, small to medium. Pileus white to ivory, aging pale smoke grey to pale vinaceous-buff to milky coffee to clay buff to drab, paler at margin, margin appendiculate. Universal veil on pileus white to ivory to pale vinaceous-buff, thin, felted, crustose or as flat patches. Lamellae white to ivory. Stipe white. Partial veil superior, white, fugacious. Universal veil at stipe base a cup forming a globose to subglobose marginate bulb. Spores amyloid, elongate to cylindrical. Universal veil of equal filamentous hyphae and inflated cells. Clamp connections absent.

Type: Australia, Western Australia: city of Melville, Murdoch University grounds, 32°4′27″S, 115°14′20″E, 28 May 2017, *E.M. Davison* 34-2017 &*P.J.N. Davison* (holo: PERTH 09178759!).

Mycobank number: MB839135.

Published sequences

Type: *ITS* (MW775291), *nuLSU* (MW775279), *ef1-α* (MW820650), *rpb2* (MW820675), *β-tubulin* (MW820660).

Description

Pileus 27–90 mm wide, up to 9 mm thick, white to ivory, aging pale cream to pale smoke grey to pale vinaceous-buff to milky coffee to clay buff to drab (5A2-4B2-5B2-6C3-6E3) with pale margin, without surface staining or bruising, initially convex becoming plane with slightly depressed centre, surface tacky when moist; margin non-striate, appendiculate. Universal veil on pileus adnate, thin, felted, crustose or breaking into flat patches over disc, white to ivory to very pale vinaceous-buff (5B2). Lamellae adnate to adnexed to free, close to subcrowded, white or ivory, 2-16 mm broad, margin concolorous, fimbriate; lamellulae plentiful, in several lengths, rounded truncate or truncate or rounded attenuate or attenuate. Stipe 26-60 mm long, 10-20 mm wide, cylindrical, or tapering upward, white, surface smooth or floccose or fibrillose below partial veil. Partial veil superior, descendent, striate above, soft to arachnoid, fugacious, white. Universal veil at stipe base a cup, forming a marginate globose to rounded bulb, $10-18 \times$ 12–22 mm, sometimes with free limb up to 5 mm high, white. Pileus and stipe context white, unchanging, stipe solid. Smell none. Spore deposit white or B (ivory).





Fig. 14. Amanita sabulosa (PERTH 09178759, holotype).

Basidiospores [220/11/11] (8.5–)10–12.5(–13) \times (4.5–)5–7 (–7.5) μ m, (L = 9.8–11.8 μ m; L' = 11.0 μ m; W = 5.2–6.4 μ m; W' = 6.0 μ m; Q = (1.39–)1.55–2.10(–2.44); Q = 1.60–2.06; Q' = 1.84), colourless, thin-walled, smooth, amyloid, elongate to cylindrical occasionally ellipsoid, contents monoguttulate or granular; apiculus sublateral, cylindric or tapered, ~1 \times 1 μ m, rounded or truncate.

Pileipellis up to 150 μm thick in old specimens, with a colourless gelatinised suprapellis up to 80 μm thick and

colourless or pale yellow subpellis, consisting of filamentous hyphae and very infrequent vascular hyphae (inflated cells not observed), filamentous hyphae 3–12 μ m wide, thick-walled, walls hyaline, gelatinising, contents colourless, radially orientated with some interweaving; vascular hyphae 2–5 μ m wide, pale yellow. Pileus context consisting of filamentous hyphae (dominant), inflated cells and very infrequent vascular hyphae, filamentous hyphae 3–40 μ m wide, with widest hyphae constricted at septa, thin-walled, colourless; inflated

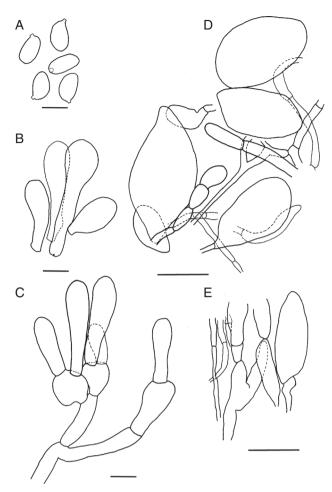


Fig. 15. Microscopic features of *Amanita sabulosa*. A. Basidiospores. B. Marginal cells. C. Basidia and sub-basidial cells. D. Universal veil from pileus, scalp section unsquashed. E. Universal veil at stipe base, inner surface of limb. Scale bars: $10~\mu m$ (A, B, C) $50~\mu m$ (D, E). (A, C, D, E PERTH 09178759, holotype; B PERTH 09178740).

cells to 350 \times 25 μ m thin-walled ventricose or clavate or cylindrical or ovoid, terminal, colourless; vascular hyphae 3–10 µm wide, occasionally branched, pale yellow. Lamella trama bilateral, divergent. Central stratum up to 20 um wide. consisting of filamentous hyphae and very infrequent vascular hyphae (inflated cells not observed), filamentous hyphae 2-15 µm wide, with widest hyphae constricted at septa, thin-walled, colourless; vascular hyphae 3-4 µm wide, some sinuous, pale yellow or colourless. Subhymenial base with angle of divergence 20-30° from central stratum with filamentous hyphae following broad curve to subhymenium, consisting of dominant filamentous hyphae, inflated cells (infrequent to frequent) and very infrequent vascular hyphae; filamentous hyphae 2-35 µm wide, widest close to subhymenium and constricted at septa, frequently branched, thin-walled, colourless; inflated cells up to $120 \times 30 \, \mu m$ when clavate or up to $120 \times 15 \mu m$ when cylindric or up to $110 \times 25 \,\mu m$ when ventricose or up to $35 \times 20 \,\mu m$ when ovoid, terminal, colourless; vascular hyphae 2-10 µm wide, occasionally branched, some sinuous, pale yellow or greenishyellow. Subhymenium with basidia arising terminally from pyriform to spherical terminal segments, becoming inflated, up to 22 µm wide. Lamella edge tissue sterile, with inflated cells infrequent to frequent, up to $45 \times 12 \,\mu m$ when clavate or up to 50×20 um when pyriform or up to 25×13 um when ovoid or up to 80×8 µm when cylindric, disarticulating, colourless. Basidia [140/7/7] (38–)40–60(–66) \times (9–)10–15(–17) μ m, thin-walled, colourless, $\sim 90\%$ 4-spored, $\sim 3\%$ 3-spored, \sim 7% 2-spored, sterigmata to 7 \times 2 μ m. Universal veil on pileus layered. Basal layer of elements with somewhat periclinal orientation, consisting of filamentous hyphae (equal or less frequent) and inflated cells (vascular hyphae not seen), filamentous hyphae 3-25 µm wide, with widest hyphae constricted at septa, colourless, some gelatinising; inflated cells up to 110 × 90 µm when ovoid or up to $120 \times 50 \mu m$ when ellipsoid or up to $70 \times 70 \mu m$ when spherical or up to $95 \times 45 \mu m$ when clavate, terminal or occasionally in chains of up to 3, disarticulating, colourless. Superficial layer very narrow, often absent, with periclinal orientation, consisting of filamentous hyphae (inflated cells and vascular hyphae not seen), 3-12 µm wide, colourless, gelatinising. Universal veil at stipe base layered. Inner layer narrow, often absent, of elements with axial orientation, consisting of filamentous hyphae (dominant to equal) and inflated cells (vascular hyphae not seen), filamentous hyphae 3-17 µm wide, some thick-walled, colourless, gelatinising; inflated cells up to 80×50 µm when ovoid or up to $150 \times 35 \,\mu m$ when clavate, terminal, colourless. Middle layer wide, consisting of filamentous hyphae (dominant to infrequent), inflated cells and very infrequent vascular hyphae, filamentous hyphae 2–12 µm wide, some thick-walled, colourless, gelatinising; inflated cells up to 140 \times 140 μm when spherical or up to $105 \times 60 \mu m$ when ovoid or up to $150 \times 50 \ \mu m$ when ellipsoidal or up to $200 \times 20 \ \mu m$ when clavate or up to 75 \times 35 μm when pyriform, terminal, colourless, disarticulating; vascular hyphae 3-6 µm wide, pale yellow or pale green. Outer layer narrow, often absent, of elements with axial orientation, consisting of dominant filamentous hyphae and frequent to infrequent inflated cells (vascular hyphae not seen), filamentous hyphae 3-20 μm wide, colourless, gelatinising; inflated cells up to $30 \times 25 \mu m$ when ovoid or up to 50×20 µm when clavate, terminal, colourless. Stipe context longitudinally acrophysalidic, consisting of frequent filamentous hyphae, dominant acrophysalides and very infrequent to frequent vascular hyphae, filamentous hyphae 2-10 µm wide, colourless; acrophysalides up to $300 \times 40 \ \mu m$ when clavate or up to $500 \times 25 \ \mu m$ when cylindric or up to 250 × 35 µm when fusiform, terminal, colourless, gelatinising; vascular hyphae 2-20 µm wide, occasionally branched, pale yellow or pale green. Ornamentation on stipe surface of filamentous hyphae (frequent or equal) and inflated cells (vascular hyphae not seen), filamentous hyphae 2-8 µm wide, colourless, disarticulating; inflated cells up to 150 \times 25 μm when clavate or up to $45 \times 25 \mu m$ when pyriform, terminal, colourless, disarticulating. Partial veil of filamentous hyphae (equal, or frequent), inflated cells and frequent vascular hyphae, filamentous hyphae 3-8 µm, wide, colourless, disarticulating; inflated cells up to 150 × 25 µm clavate,

colourless, disarticulating; vascular hyphae 2–8 µm wide occasionally branched, colourless or pale yellow or pale greenish yellow. Clamp connections not observed.

Habit, habitat and distribution

Singly or gregarious in sand, in native vegetation; nearby plants include *Allocasuarina fraseriana*, *Brachyloma preissii*, *Corymbia calophylla*, *Eucalyptus marginata*, *Jacksonia furcellata* and *Melaleuca lanceolata*. Occurs in the Swan Coastal Plain Perth SWA02 and IBRA subregion and the Murray—Darling Depression Murray Mallee MDD02 IBRA subregion (Department of the Environment 2013). Fruiting period is May to July.

Etymology

From the Latin *sabulosus* meaning 'sandy', referring to the sandy soil where it occurs.

Notes

Figure 1 shows *A. sabulosa* is a member of section *Arenariae*. The pileus is, in appearance, similar to that of *A. pupatju* and the spores are of similar shape (*A. sabulosa* [220/11/11] $\mathbf{L} = 9.8{-}11.8~\mu m$, $\mathbf{W} = 5.2{-}6.4~\mu m$, $\mathbf{Q} = 1.60{-}2.06$; *A. pupatju* [200/10/8] $\mathbf{L} = 10.6{-}13.6~\mu m$, $\mathbf{W} = 5.2{-}6.5~\mu m$, $\mathbf{Q} = 1.80{-}2.12$); however, these two species differ in the shape of the stipe base, which is conic to tapered in *A. pupatju*, whereas *A. sabulosa* has a marginate bulb (Fig. 12, 14). The nuLSU, $efl{-}\alpha$, β -tubulin sequences show that these two species differ (Fig. 1, 2A, C), and *ITS* sequences differ by 14.6–18.0% (Table 4).

Amanita sabulosa has spores of similar shape to those of A. lesueurii ([200/10/5] $L=11.1-12.4~\mu m$, $W=5.5-6.2~\mu m$, Q=1.90-2.26), but lacks the felted or floccose universal veil on the pileus, which becomes vinaceous-buff to grey with age (Davison et al. 2013). The spores of A. sabulosa are wider than those of A. wadulawitu ([859/21/21] $L=10.2-12.5~\mu m$, $W=4.4-5.2~\mu m$, Q=2.21-2.55) and it has a universal veil, which forms a marginate bulb, not a bulb that is initially turbinate and which elongates with age (McGurk et al. 2016).

There are other species described from Australia that have amyloid spores, a felted universal veil and no clamp connections, that may fall with section Arenariae. Amanita sublutea (Cleland) E.-J.Gilbert is a poorly known species from South Australia. From the descriptions given by Cleland (1931), Reid (1980), Bas (1969) and Grgurinovic (1997), A. sabulosa has a paler pileus, wider stipe, shorter and narrower spores (A. sublutea: [20/2] (11–)11.5–13(–13.5) \times 6.5–7.5(–8) μ m (Bas 1969), [50/2] 11–13.8 \times 5.8–7.7 μ m (Grgurinovic 1997); A. sabulosa [220/11/11] (8.5–)10–12.5 $(-13) \times (4.5-)5-7(-7.5)$ µm), the subhymenium is inflated rather than inflated ramose and its cells are not coralloid. Some of these differences may be the result of environmental conditions; more collections of A. sublutea from the type locality, or sequences from the holotype, would clarify whether these are two different species.

Amanita sabulosa differs from A. gracilenta A.E. Wood by its larger size, more robust form, marginate, not slightly swollen, bulb, presence of a partial veil, shorter and

narrower spores (*A. gracilenta* $8.1-9.9(-11.0) \times 4.5-6.1$ (-6.6) µm; *A. sabulosa* [200/10/10] ($9-10-12.5(-13) \times (4.5-)5-7(-7.5)$ µm), and larger inflated cells in the universal veil on the pileus (Wood 1997). It differs from *A. clelandii* and *A. variabilis*, two poorly known species from South Australia, by its larger size and different-shaped spores (Reid 1980). On this basis, *A. sabulosa* is described as a new species.

Specimens examined

WESTERN AUSTRALIA. City of Melville, E.M. Davison 10-1995 & P.J.N. Davison (PERTH 09269452); loc. id., E.M. Davison 11-1995 & P.J.N. Davison, (PERTH 09269487); loc. id., E.M. Davison 11-2002 & P.J.N. Davison (PERTH 09178694); loc. id., E.M. Davison 23-2003 & P.J.N. Davison (PERTH 09178708); loc. id., E.M. Davison 15-2014 & P.J.N. Davison (PERTH 09178716); loc. id., E.M. Davison 17-2014 & P.J.N. Davison (PERTH 09178724); loc. id., E.M. Davison 15-2015 & P.J.N. Davison (PERTH 09178732); loc. id., E.M. Davison 41-2016 & P.J.N. Davison (PERTH 09178740); loc. id., E.M. Davison 35-2017 & P.J.N. Davison (PERTH 09178767); loc. id., E.M. Davison 57-2017 & P.J.N. Davison (PERTH 09178775).

SOUTH AUSTRALIA. Monarto Conservation Park, P.S. Catcheside 2509 & D.E.A. Catchside (AD-C54481).

Amanita griselloides D.A.Reid, Victorian Naturalist 95(2): 47 (1978)

(Fig. 16, 17)

Basidiomata agaricoid, small to medium. Pileus pale grey to grey to pale brown to brown, paler at margin, margin appendiculate. Universal veil on pileus white, becoming greyish-brown to brown, felted to floccose, as one or several patches. Lamellae white. Stipe white with ovoid to conic bulb that elongates with age. Partial veil superior to median, white, fugacious. Universal veil at top of bulb very inconspicuous or absent. Spores amyloid, ellipsoid to elongate. Universal veil on pileus of dominant inflated cells with brown or greyish-brown contents. Clamp connections absent.

Type: AUSTRALIA. Western Australia: 2 km north of Bow River on Highway 1, near Walpole, 34°37′S, 116°58′E, 13 May 1976, *D.A & D.G Reid, R. Hilton, N. Brittan* (holo: K(M) 236386!; iso: DAR 32037!).

Mycobank number: MB308554.

Description

Pileus 33–65 mm wide, up to 10 mm thick, silvery-grey to smoke grey to silvery-beige to cinnamon to pale brown to brown, margin paler, without surface staining or bruising, initially convex, becoming plane with decurved margin and slightly depressed centre, surface slightly tacky when moist shiny when dry; margin non-striate, slightly appendiculate when young. Universal veil on pileus adnate, felted to floccose, as one or several patches in centre of disc, white to light brown to greyish-brown to brown. Lamellae adnexed to adnate, close to subcrowded, white to pale buff (5B2), 6–10 mm broad, margin concolorous, fimbriate; lamellulae frequent to infrequent in several lengths, shortest truncate or rounded truncate, longest attenuate or subattenuate. Stipe 20–48 mm long, 8–27 mm wide, cylindrical or tapering



Fig. 16. Amanita griselloides (PERTH 09004831).

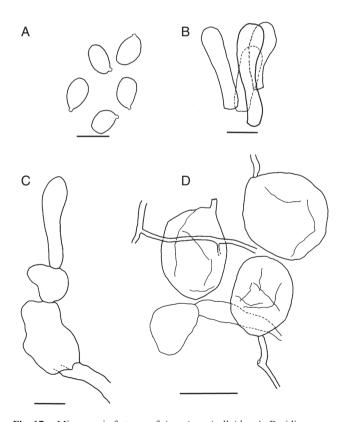


Fig. 17. Microscopic features of *Amanita griselloides*. A. Basidiospores. B. Marginal cells. C. Basidium and sub-basidial cell. D. Universal veil from pileus, squashed. Scale bars: 10 μ m (A, B, C) 50 μ m (D). (K(M) 236386, holotype).

upwards, white or cream, surface at top of stipe powdery, below smooth or minutely floccose or fibrillose. Partial veil superior to median, descendent, striate above, fugacious or not observed, white or pale buff. Bulb $25-30\times 8-32$ mm, ovoid or conic becoming tapered, white or pale buff. Remains of universal veil at top of bulb not apparent or a soft, floccose rim at top of bulb, white. Pileus and stipe context white, unchanging, stipe solid, becoming hollow. Smell not distinctive or unpleasant. Spore deposit white.

Basidiospores [180/9/8] (8–)8.5–12(–12.5) \times (5–)5.5–7 μ m, (L = 9.2–11.3 μ m; L' = 10.0 μ m; W = 5.9–6.6 μ m; W' = 6.2 μ m; Q = (1.29–)1.38–1.83(–2.00); Q = 1.51–1.76; Q' = 1.61), colourless, thin-walled, smooth, amyloid, ellipsoid to elongate, contents monoguttulate or granular; apiculus sublateral, cylindric or tapered, ~1 \times 1 μ m, truncate or rounded.

Pileipellis poorly developed in young specimens, up to 200 µm thick in old specimens, with a colourless gelatinised suprapellis up to 150 µm thick, and yellow or brown subpellis, consisting of filamentous hyphae and frequent to infrequent vascular hyphae (inflated cells not observed), filamentous hyphae 2-12 µm wide, thick-walled, walls hyaline, gelatinising, contents colourless to yellow to brown, radially orientated; vascular hyphae 2-10 µm wide, occasionally branched, pale yellow or yellowish-brown. Pileus context consisting of filamentous hyphae (dominant or equal), inflated cells and frequent to infrequent vascular hyphae, filamentous hyphae 3-25 µm wide, with widest hyphae constricted at septa, thin-walled, colourless; inflated cells up to 300 \times 35 μ m, thin-walled, some gelatinising, clavate or ventricose or ovoid or cylindrical, colourless; vascular hyphae 3-15 µm wide, occasionally branched, pale yellow. Lamella

trama bilateral, divergent. Central stratum up to 30 µm wide, consisting of filamentous hyphae (inflated cells and vascular hyphae not observed), filamentous hyphae 4-6 µm wide, colourless. Subhymenial base with angle of divergence ~30° from central stratum, with filamentous hyphae following smooth broad curve to subhymenium, consisting of dominant filamentous hyphae, inflated cells (infrequent or not seen; vascular hyphae not seen), filamentous hyphae 3-15 µm wide, widest close to subhymenium and constricted at septa, frequently branched, thin-walled, colourless; inflated cells up to 70 × 20 µm ventricose or cylindric, terminal, colourless. Subhymenium with basidia arising terminally from barely inflated to pyriform terminal segments up to 18 µm wide. Lamella edge tissue sterile, with inflated cells infrequent to frequent, to $20-35 \times 10-18 \ \mu m$ clavate or pyriform or ovoid, colourless, disarticulating. Basidia [140/7/7] (31–)35–59(–66) \times (8–)9–13(–15) μ m, thin-walled, colourless, $\sim 86\%$ 4-spored, $\sim 8\%$ 3-spored, \sim 6% 2-spored, sterigmata up to 8 \times 2 μ m. Universal veil on pileus not layered, elements with no dominant orientation, consisting of frequent to dominant filamentous hyphae, dominant to frequent inflated cells and very infrequent to frequent vascular hyphae, filamentous hyphae 3-15 µm wide, colourless or pale brown or pale greyish-brown, gelatinising; inflated cell up to $85 \times 70 \,\mu m$ when ovoid or up to $70 \times 70 \,\mu m$ when spherical or up to $90 \times 50 \,\mu m$ when ellipsoidal or up to $110 \times 25 \mu m$ when ventricose or up to $110 \times 45 \,\mu m$ when clavate, terminal, occasionally in chains of up to 3 cells, colourless or grey or greyish-brown or brown, gelatinising; vascular hyphae 3-12 µm wide, occasionally branched, brownish-yellow. Universal veil on stipe base often absent, not layered, with no dominant orientation, consisting of dominant filamentous hyphae, frequent inflated cells and very infrequent vascular hyphae, filamentous hyphae 4–10 µm wide, thick-walled, colourless or pale grey, gelatinising; inflated cells up to $110 \times 20 \mu m$ when clavate or up to 110×40 µm when pyriform, very pale grey; vascular hyphae 5 µm wide, pale yellow. Stipe context longitudinally acrophysalidic, consisting of frequent to equal filamentous hyphae, acrophysalides and infrequent to frequent vascular hyphae, filamentous hyphae 2-10 µm wide, colourless; acrophysalides up to 190 × 35 µm, clavate or cylindric, terminal, colourless; vascular hyphae 3-35 µm wide, occasionally branched, yellowish-brown. Partial veil not examined. Clamp connections not observed.

Habit, habitat and distribution

Gregarious in sand. In native vegetation; nearby plants include *Agonis hypericifolia*, *Allocasuarina fraseriana*, *Eucalyptus diversicolor* and *E. marginata*. Occurs in the Southern Jarrah Forest JAF02, Warren WAR01, and Recherche ESP02 IBRA subregions (Department of the Environment 2013). Fruiting period is April to July.

Notes

Reid (1980) described the universal veil on the pileus of *A. griselloides* as layered, with a grey felty floccose layer

of inflated cells with brown contents intermixed with thin-walled, hyaline hyphae, covered by a white arachnoid superficial layer formed entirely of thin-walled, prostrate hyphae. He commented that the structures of the universal veil on the pileus of an additional collection (DAR 32036) were exactly as in the type. A layered universal veil in which there is a superficial layer of mainly filamentous hyphae occurs in some members of subgenus *Amanitina* (Bas 1969, pp. 307–308). However, in the type K(M) 236386, this superficial layer has phialides, indicating it is a saprophyte growing on the basidiome. This superficial layer was not seen in DAR 32036. It has not been seen in the other collections of *A. griselloides*.

Specimens examined

WESTERN AUSTRALIA. City of Albany, *G. Byrne 5790* (PERTH 09175938); *loc. id.*, *G. Byrne 626628* (PERTH 09004831); *loc. id.*, *D.A. & D.G. Reid, R Hilton & N. Brittan s.n.* (DAR 32036); shire of Esperance, *E.M. Davison* 2–1990 & *P.J.N. Davison* (PERTH 03096866); *loc. id.*, *E.M. Davison* 5-1990 & *P.J.N. Davison* (PERTH 03096858); shire of Denmark, *K. Syme 5/91* (PERTH 04988442); city of Albany, *K. Syme 400/92* (PERTH 05254299).

Amanita peltigera D.A.Reid, Victorian Naturalist 95(2): 49 (1978)

Type: AUSTRALIA. South Australia: Stirling West, 35°00′S, 138°43′E, Mar. 1976, J. Randles s.n. (holo: K(M) 236385!).

Mycobank number: MB308577.

Amanita dumosorum D.A.Reid, Victorian Naturalist 95(2): 47 (1978), Type: AUSTRALIA. Western Australia: city of Albany, Two Peoples Bay Nature Reserve, 34°58′S, 118°10′E, 12 May 1976, D.A. & D. G. Reid, R. Hilton & N. Brittan (holo: K(M) 236387! iso: DAR 32034!), syn. nov.

Mycobank number: MB308550.

Notes

As mentioned in the Introduction, Reid (1980) considered A. dumosorum and A. peltigera to be similar, apart from the presence of a partial veil and the absence of a saccate volva (Fig. 18). However, a study of collections of A. peltigera from SA and WA showed variation in the presence of the partial veil and the development and thickness of the universal veil at the top of the bulb (Davison et al. 2020). Reid (1980) also stated that the structure of the universal veil on the pileus was entirely hyphal; however, we have noted infrequent, large, inflated cells in this tissue (Fig. 19). Basidiospores are amyloid and of similar size: A. peltigera [499/25/25] $(6.5-)8-10.5(-11.5) \times$ (5.5-)6-8(-9) µm, (L = 8.0-10.0 µm; L' = 9.1 µm; $\mathbf{W} = 6.3-7.7 \ \mu\text{m}; \ \mathbf{W}' = 7.1 \ \mu\text{m}; \ \mathbf{Q} = (1.06-)1.14-1.43$ (-1.62); $\mathbf{Q} = 1.20 - 1.38$; $\mathbf{Q}' = 1.29$), A. dumosorum: [40/1/1] $7-10(-10.5) \times 5.5-7.5 \ \mu\text{m}, \ (\mathbf{L}' = 9.0 \ \mu\text{m}; \ \mathbf{W}' = 6.5 \ \mu\text{m};$ Q = (1.17-)1.20-1.54; Q' = 1.38), and their microscopic features are similar (Fig. 19). On this basis, A. dumosorum is synonymised with A. peltigera.

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 $\textbf{Fig. 18.} \quad \textit{Amanita dumosorum} \, (K(M) \, 236387, holotype). \, Images: J. \, Percy-Bower, \, Western \, Australian \, Herbarium.$

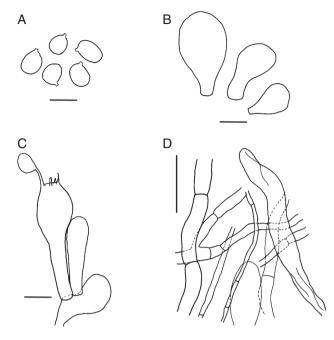


Fig. 19. Microscopic features of *Amanita dumosorum*. A. Basidiospores. B. Marginal cells. C. Basidia and sub-basidial cell. D. Universal veil from pileus, scalp section gently squashed. Scale bars: $10~\mu m$ (A, B, C) $50~\mu m$ (D). (K(M) 236387, holotype).

Key

1.	Basidiome agaricoid2
	Basidiome secotioid
2.	Pileus and universal veil on pileus white or pale3
	Pileus aging grey, brown or greyish-brown
3.	Bulb initially turbinate, elongating with age
	Bulb conic or marginate
4.	Universal veil on pileus white, unchanging, spores
	cylindrical
	Universal veil on pileus becoming pale brown to grey, spores elongate to
	cylindrical
5.	Bulb conic
	Bulb marginate
6.	Universal veil on pileus white, spores broadly ellipsoid to
	ellipsoid
	Universal veil on pileus grey to brown, spores ellipsoid to elongate
	7
7.	Pileus aging brown, universal veil on pileus felted to
	floccose
	Pileus aging grey, universal veil on pileus felted, breaking into
	aerolate patches
8.	Clamp connections present Section Amarendiae (not dealt with here)
	Clamp connections absent
9.	Basidiome cylindric
	Basidiome capitate
10.	Spores subglobose to broadly ellipsoid ($\mathbf{Q} = 1.11-1.27$)
	A. pseudoarenaria
	Spores ellipsoid to cylindric
11.	Spores ellipsoid to elongate (Q = 1.54-1.70), south coast
	distribution
	Spores elongate to cylindrical (Q = 1.85–2.13) A. arenarioides

Discussion

The recent revision of Amanitaceae by Cui et al. (2018), by using concatenated datasets of nuLSU, β -tubulin, ef1- α and rpb2 gene regions, provides a phylogeny for Amanita species with amyloid spores. Our work, using only the nuLSU gene region, shows that section Arenariae is sister to sections Phalloideae, Strobiliformes and Validae, with good bootstrap support (Fig. 1). Within section Arenariae, there is a well-supported clade containing the secotioid species arenaria, A. arenarioides, A. compacta and agaricoid pseudoarenaria. and the species A. griselloides, A. pupatju and A. sabulosa (Fig. 2B-D). There are insufficient sequences to resolve their relationships with A. lesueurii, A. peltigera, A. wadjukiorum and A. wadulawitu. the other known members of this section.

The multicopy ITS region is the DNA barcode marker for fungi (Schoch et al. 2012). We have found that cloning is necessary to consistently obtain high-quality sequences (McGurk 2013; McGurk et al. 2016; Davison and Giustiniano 2020; Davison et al. 2017a, 2017b, 2020). However, cloning has shown inconsistent utility of this region for delineating species from different sections of Amanita from southern Australia (Davison et al. 2017a, 2020). In section Phalloideae, ITS sequences varied by up to 1.42% among collections of the same species, and by 1.42-1.60% between different species which morphologically distinct and geographically separated (Davison et al. 2017a); however, in section Arenariae, amplicons from the same individual vary from 0.0 to 9.9% (Tables 3, 4; Davison et al. 2020). There were no changes to the methodology and processing of samples from sections Phalloideae and Arenariae. Examination of the ITS region showed consistent insertions and deletions among different species, and some consistent transitions and transversions among collections of the same species. We have assumed that these represent real differences among clones, not being PCR-induced errors; no chimeras were detected by UCHIME. We were not able to determine whether these differences represent past hybridisation events (Hughes et al. 2015, 2018).

Section Arenariae

It is likely that there are many more described agaricoid species in Australia that are unrecognised members of this section. These would be expected to have a slightly appendiculate pileus margin, and inflated sub-basidial cell; possible examples are *A. gracilenta*, *A. albidannulata* A.E. Wood, and *A. annulalbida* A.E. Wood.

Data availability statement

The data that support this study are available in the article.

Conflicts of interest

The authors declare that they have no conflicts of interest.

Declaration of funding

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