

Supplementary material

Phytoplankton–zooplankton relationships based on phytoplankton functional groups in two tropical reservoirs

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Table S1. Morphological and physiological characteristics (toxicity), representative taxa for each morpho-functional group, and the degree of susceptibility to predation by zooplankton

Adapted from Colina *et al.* (2016)

MBFG	Characteristics	Representative taxon	Toxic potential	Susceptibility to predation
I	Small organisms occupying large areas	<i>Merismopedia tenuissima</i> , <i>Planktosphaeria gelatinosa</i>	0	High
II	Small flagellates with siliceous structures	<i>Mallomonas caudata</i>	0	Low
III	Broad filaments with gas-vacuoles	<i>Cylindrospermopsis raciborskii</i> , <i>Planktothrix agardhii</i>	1	Low
IV	Organisms of medium size without specialised structures	<i>Cosmarium</i> sp., <i>Staurastrum</i> sp., <i>Geitlerinema amphibium</i>	0	High
V	Medium to large single-celled flagellates	<i>Rhodomonas lacustris</i> , <i>Peridinium</i> sp.	1	Medium
VI	Organisms without flagella with siliceous exoskeleton	<i>Aulacoseira granulata</i> , <i>Thalassiosira</i> sp.	0	Medium
VII	Large mucilaginous colonies	<i>Microcystis aeruginosa</i> , <i>Botryococcus braunii</i>	1	Low

Table S2. Taxonomic composition of the phytoplankton community by morphology-based functional groups (MBFG) in the mesotrophic reservoir

MBFG	Species identified
I	<i>Ankistrodesmus fusiformis</i> , <i>Merismopedia tenuissima</i> , <i>Monoraphidium griffithii</i> and <i>Planktosphaeria gelatinosa</i> .
II	<i>Mallomonas caudata</i> .
III	<i>Cylindrospermopsis raciborskii</i> , <i>Dolichospermum</i> sp. and <i>Planktothrix agardhii</i> .
IV	<i>Closterium</i> sp., <i>Closterium incurvum</i> , <i>Coelastrum indicum</i> , <i>Coenocystis</i> sp., <i>Cosmarium commissurale</i> , <i>C. margaritatum</i> , <i>Desmodesmus quadricauda</i> , <i>Dictyosphaerium elegans</i> , <i>Euastrum abruptum</i> , <i>Eutetramorus nygaardii</i> , <i>E. planctonicus</i> , <i>Geitlerinema amphibium</i> , <i>Kirchneriella obesa</i> , <i>Pediastrum tetras</i> , <i>Pseudanabaena</i> sp., <i>Quadrigula closterioides</i> , <i>Scenedesmus acuminatus</i> , <i>Sphaeroscystis schroeteri</i> , <i>Staurastrum curvimarginatum</i> , <i>S. cuspidatus</i> , <i>S. dilatatum</i> , <i>S. leptocladum</i> , <i>S. orbiculare</i> , <i>S. tetracerum</i> , <i>S. trifidum</i> , <i>Staurastrum</i> sp., <i>Tetraedron gracile</i> , <i>T. mediocris</i> and <i>T. trigonum</i> .
V	<i>Chroomonas</i> sp., <i>Cryptomonas</i> sp., <i>Euglena</i> sp., <i>Peridinium</i> sp., <i>Rhodomonas lacustris</i> , <i>Trachelomonas hispida</i> and <i>T. volvocina</i> .
VI	<i>Aulacoseira granulata</i> , <i>Cylindrotheca closterium</i> , <i>Cymbella</i> sp., <i>Eunotia</i> sp., <i>Navicula</i> sp., <i>Surirella</i> sp., <i>Surirella tenera</i> , <i>Thalassiosira</i> sp. and <i>Ulnaria ulna</i> .
VII	<i>Aphanocapsa</i> sp. and <i>Botryococcus braunii</i> .

Table S3. Taxonomic composition of the phytoplankton community by morphology-based functional groups (MBFG) in the supereutrophic reservoir

MBFG	Species identified
I	<i>Merismopedia tenuissima</i> , <i>Monoraphidium griffithii</i> and <i>Planktosphaeria gelatinosa</i> .
II	<i>Mallomonas caudata</i> .
III	<i>Anabaena</i> sp., <i>Cylindrospermopsis raciborskii</i> , <i>Dolichospermum viguieri</i> , <i>Planktothrix agardhii</i> and <i>Sphaerospermopsis aphanizomenoides</i> .
IV	<i>Actinastrum hantzschii</i> , <i>Closterium</i> sp., <i>Coelastrum indicum</i> , <i>Dictyosphaerium elegans</i> , <i>D. pulchellum</i> , <i>Geitlerinema amphibium</i> , <i>Kirchneriella obesa</i> , <i>Micractinium pusillum</i> , <i>M. quadrisetum</i> , <i>Pseudanabaena</i> sp. and <i>Scenedesmus acuminatus</i> .
V	<i>Chroomonas</i> sp., <i>Cryptomonas</i> sp., <i>Euglena</i> sp., <i>Peridinium</i> sp., <i>Rhodomonas lacustris</i> , <i>Trachelomonas hispida</i> and <i>T. volvocina</i> .
VI	<i>Aulacoseira granulata</i> , <i>A. granulata</i> var. <i>angustissima</i> , <i>Cylindrotheca closterium</i> , <i>Cyclotella meneghiniana</i> , <i>Eunotia</i> sp., <i>Gomphonema</i> sp., <i>Navicula</i> sp. and <i>Ulnaria ulna</i> .
VII	<i>Aphanocapsa</i> sp., <i>Botryococcus braunii</i> , <i>Microcystis aeruginosa</i> , <i>M. brasilensis</i> , <i>M. panniformis</i> and <i>Woronichinia karelica</i> .

Table S4. Statistical analysis (one way ANOVA) of the growth rate of morphology-based functional groups (MBFG) in *in situ* experiments

Bold numbers represents that there was a significant difference

	Mesotrophic			Supereutrophic		
	d.f.	F	P	d.f.	F	P
Group I	2	2.082	0.206	2	4.169	0.073
Group II	2	4.408	0.057	2	2.049	0.209
Group III	2	1.942	0.224	2	0.094	0.912
Group IV	2	7.513	0.023	2	0.754	0.51
Group V	2	3.103	0.119	2	16.01	0.003
Group VI	2	1.204	0.364	2	9.694	0.013
Group VII	2	1.886	0.231	2	2.46	0.166

Table S5. Statistical analysis (Student *t*-test unpaired) of the ingestion rate of micro and mesozooplankton on morphology-based functional groups (MBFG) in *in situ* experiments

	Mesotrophic			Supereutrophic		
	d.f.	<i>t</i>	<i>P</i>	d.f.	<i>t</i>	<i>P</i>
Group I	3.769	-1.919	0.131	2.436	-0.781	0.503
Group II	2.377	0.688	0.552	4	0	1.000
Group III	2.717	0.535	0.632	3.668	-2.486	0.073
Group IV	3.864	-2.541	0.066	3.752	-0.037	0.972
Group V	3.544	-0.854	0.446	2.768	-0.733	0.520
Group VI	3.634	-1.284	0.274	3.252	-2.437	0.086
Group VII	3.999	1.530	0.200	3.445	0.172	0.873

Reference

Colina, M., Calliari, D., Carballo, C., and Kruk, C. (2016). A trait-based approach to summarize zooplankton–phytoplankton interactions in freshwaters. *Hydrobiologia* **767**, 221–233. [doi:10.1007/s10750-015-2503-y](https://doi.org/10.1007/s10750-015-2503-y)