Accessory Publication

Relative salinity tolerance of macroinvertebrates from the Barwon River, Victoria, Australia

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Abstract. Salinity levels are rising in many freshwater environments, yet there are few direct measurements of salinity tolerance of organisms likely to be salt sensitive. The relative salinity tolerance to artificial seawater of macroinvertebrates from the Barwon River in Victoria, Australia, was assessed by measuring the 72-h lethal concentrations required to kill 50% of individuals (LC_{50}). LC_{50} values ranged from an electrical conductivity of 5.5 to 76 mS cm⁻¹ (mean 31 mS cm⁻¹, n = 57) and followed a log-normal distribution. The most salt-sensitive groups tested were Baetidae (LC_{50} value range: 5.5–6.2 mS cm⁻¹), Chironomidae (10 mS cm⁻¹) and several softbodied non-arthropods (Oligochaeta, Gastropoda, Nematomorpha, Tricladida and Hirudinea; 9–14 mS cm⁻¹). Other groups, from least to most tolerant, were non-baetid Ephemeroptera (>12.6–15 mS cm⁻¹), Plecoptera (>12.6–>20 mS cm⁻¹), Trichoptera (9–>26 mS cm⁻¹), Corixidae (18–26 mS cm⁻¹), non-corixid Hemiptera (33–44 mS cm⁻¹), Coleoptera (19–54 mS cm⁻¹), Hydracarina (39 mS cm⁻¹) and Odonata (30–55 mS cm⁻¹), and macrocrustaceans (Decapoda, Isopoda and Amphipoda; 38–76 mS cm⁻¹).

Lethal salinity tolerance and selected field distributions of freshwater macroinvertebrate taxa tested in the present study

Where results were available for a number of methods, water temperatures, etc., the information given is for the method most comparable with the present study. The tolerances were established in artificial seawater (ASW), seawater (SW), saline lake water (SLW), oil brine (OB), estuary water (EW), evaporated river water (ERW), food salt (FS), sodium chloride (NaCl), sodium sulfate (Na₂SO₄) or, where not given, it is unspecified in the original publication. ASW, SW, SLW, EW and OB were diluted were appropriate. The time of the test is given in hours or days.

Гаха	Location	Measure and salt source	Time	Tolerance (mS cm ⁻¹)	Reference
Gastropoda			0.61	0.5	
Physa	USA USA	LC_{50} in OB	96 h 96 h	8.5 7.1	1
Physa Burnupia sternchorias (Ancylidae)	South Africa	LC_{50} in NaCl LC_{50} in Na ₂ SO ₄	96 h 96 h	7.1	1
Potamopyrgus antipodarum	New Zealand	Remains out of its			3
Hydrobiidae)	Australia	LC in SW	96 h	31	4
<i>Posticobia</i> sp. (Hydrobiidae) <i>Coxiella</i> (Pomatiopsidae)	Australia	LC ₅₀ in SW Remains out of its she			4 5
Dligochaeta Eight Tubificidae, one Lumbriculidae	North America	Range in LC ₅₀ values	96 h	9.9–14	6, 7
Hirudinea					
All Hirudinea	Canada	Maximum field distribution of 3 above	8		
Neophelopsis obscura (Erpodellidae)	Canada	Die within 2 h at 1.2 but survi	8		
Actinobdella inequiannulata (Glossiphoniidae)	Not given	Only surviv	9		
Baetidae (Ephemeroptera)					
Cloen crassi	South Africa	Tolerance in SW		9.5-14	10
Cloen africanum	South Africa	Tolerance in SW		4.6–9.3	10
Jnidentified Baetidae	USA	LC ₅₀ in OB	96 h	11	1
Aix of Baetidae	South Africa	LC ₅₀ in NaCl	96 h	5.0-10	2
Aix of Baetidae	South Africa	LC_{50} in Na_2SO_4	96 h	10	2
<i>Fricotythus</i> sp.	South Africa	LC_{50} in NaCl	96 h	1.0	11
Tricotythus sp. Tricotythus tinctus	South Africa South Africa	LC_{50} in Na_2SO_4 Mean LC ₂₂ in $NaCl$	96 h 96 h	4.0-8.0 2.9	11 12
ricotythus tinctus ricotythus tinctus	South Africa	Mean LC ₅₀ in NaCl Mean LC ₅₀ in Na ₂ SO ₄	96 h 96 h	2.9	12
froptilum sudafricanum	South Africa	LC_{50} in Na_2SO_4	96 h	2.8	12
denophlebia auriculata	South Africa	LC_{50} in Na ₂ SO ₄ LC ₅₀ in Na ₂ SO ₄	96 h	5.3	12
horoterpes sp.	South Africa	LC_{50} in NaCl	96 h	2.9	12
aetis rhodani	UK	Survived 4 h in SW a			13
chithrogena semicolorata	UK	Survived 4 h in SW at 12 but not at 23 Survived 4 h in SW at 12 but not at 23			13
Other Ephemeroptera					
Aix of Leptophlebiidae and Caenidae	Australia	LC ₅₀ in FS	96 h	20	14
<i>Iexagenia</i> (Ephemeridae) All Ephemeroptera	USA Australia	LC ₅₀ in OB Maximum field d	96 h listribution 5.7	14 7	1 15
Plecoptera					
Dinocras cephalotes and Perla bipunctata (Perlidae)	UK	Both survived 6 h in S	13		
Small unidentified Nemouridae	UK	Could not survive	13		
All Plecoptera	Australia	Maximum field distribution 4.2			15
All Plecoptera	Spain	Maximum field d	listribution 8.4	1	16
Paragnetina media (Perlidae)	Canada	LC50 in NaCl	72 h	16–21	17
Trichoptera Cheumatopsyche modica, C. sp. 1 and Asmicridea edwardsii	Australia	Increased mortality in laboratory tests	96 h	1.6–3.2	18
Hydropsychidae) Phryganea grandis and P. pagetana	Norway	LC ₅₀	96 h	>27	19
Phryganeidae) Limnephilus marmoratus	Norway	LC_{50}	96 h	Less tolerant	19
Limnephilidae) Limnephilus affinis	UK	Survived several	monthe in 24	than above	20
Limnephilus affinis L. stigma and Anabolia nervosa Limnephilidae)	UK	Died with 3 day	20 21		
Corixidae (Hemiptera)					
All Corixidae	Australia	Maximum field d			22
igara australis and Agraptocorixa itifrons	Australia	Did not survive in laboratory much beyond 18			20
Micronecta gracilis	Australia	Unaffected	up to 7.9		18
Other Hemiptera Anisops barbata (Notonectidae)	India	Survived 76 h	in SW at 10		23
	mula		23		
Sphaerodema rusticum	India	Survived 72 h	in SW at 25		23
Sphaerodema rusticum Belostomatidae) Ranatra elogata (Nepidae)	India India	Survived 72 h Survived 48 h			23

Coleoptera						
<i>Cybister tripunctalus, C. cognatus, C. sugillatus</i> (Dytisicidae)	India	Survived between 66 ar	23			
<i>C. tripunctatus</i>	India	Survived 65 h in SW at 33				
Diaptomus clavipes (Diaptomidae)	USA	LC ₅₀ in OB	96 h	8.7	23 1	
Diaptomus clavipes	USA	LC_{50} in NaCl	96 h	5.6	1	
Adult Elmidae	Australia	Mortal above	96 h	4.8	18	
	Tuotiunu		<i>y</i> 0 H		10	
Odonata						
Bradynopyga geminata (Libellulidae)	India	Survived 96 h i		23		
Tramea virgina (Libellulidae)	India	Survived 96 h in SW at 19			23	
Idictinogomphus rapax (Gomphidae)	India	Survived 86 h in SW at 23			23	
Hemianax ephippiger (Aeshnidae)	India	Survived 90 h in SW at 19			23	
Unidentified Libellulidae	USA	LC ₅₀ in oil brine	96 h	20	1	
Unidentified Libellulidae	USA	LC50 in NaCl	96 h	21	1	
Unidentified Coenagrionidae	USA	LC_{50} in oil brine	96 h	20	1	
C C						
Decapoda						
Paratya australiensis	Australia	LC ₅₀ in EW	72 h	29	24	
P. australiensis	Australia	LC ₅₀ in SW	96 h	34	25	
P. australiensis	Australia	LC ₅₀ in SW	96 h	35	4	
P. australiensis	Australia	LC ₅₀ in ASW	96 h	6.0	26	
P. australiensis	Australia	LC ₅₀ in SW	96 h	30	27	
Caridina nilotica (Atyidae)	South Africa	LC50 to mix of NaCl and	96 h	4.4	12	
		Na_2SO_4				
Amarinus lacustris	Australia	LC _{3.5} in SW	96 h	61	4	
A. lacustris	Australia	LC50 slow acclimatization in ERW	Weeks	48	28	
Cherax quadricarinatus (Parastacidae)	Australia	LC ₃₀ in SW	21 days	32	29	
<i>C. destructor</i>	Australia	LC_{50} juvenile and adult in SW	21 days 96 h	34, 40	30	
Macrobrachium australiense	Australia	LC ₅₀ in SW	96 h	48	4	
(Palaemonidae)	7 tustrana	LC50 III 5 W	90 II	40	-	
M. potiuna	Brazil	LC ₅₀ in SW	48 h	19	31	
<i>Cambarus</i> sp. (Cambaridae)	USA	LC_{50} in OB	96 h	24	1	
Cambarus sp. (Cambardad)	USA	LC_{50} in NaCl	96 h	23	1	
Cambaras sp.	00/1	2030 m Huer	70 H	25	1	
Amphipoda						
Austrochiltonia australis	Australia	LC ₅₀ in SW	96 h	39	4	
A. subtenuis	Australia	LC ₅₀ in SW	96 h	42	4	
Hyalella azteca (Talitridae)	USA	LC ₅₀ in OB	96 h	14	1	
H. azteca	USA	LC ₅₀ in SLW	96 h	25	32	
Gammarus fossarum (Gammaridae)	Netherlands Survived half as long as specimens in controls in SW at 4.0					
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1, Clemens and Jones (1954); 2, Palmer *et al.* (1996); 3, Winterbourn (1970); 4, Williams (1984); LC₅₀ values calculated from raw data; 5, Williams and Mellor (1991); 6, Chapman and Brinkhurst (1980); 7, Chapman *et al.* (1982); 8, Scudder and Mann (1968); 9, Sawyer (1986); 10, Forbes and Allanson 1970, 11, Goestsch and Palmer (1997); 12, Palmer and Scherman (2000); 13, Williams and Williams (1998); 14, Kefford and Robley (1996); 15, Kefford (1998); 16, Prenda and Gallardo-Mayneco (1999); 17, Kapoor (1979); 18, Metzeling *et al.* (1995); 19, Haage (1969); 20, Sutcliffe (1960); 21, Sutcliffe (1961); 22, Knowles and Williams (1973); 23, Shirgur and Kewalramani (1973); 24, Walker (1972); 25, Hargraves (1975); 26, Bacher and Garnham (1992); 27, Walsh (1994); 28, Walker (1969); 29, Jones (1990); 30, Mills and Geddes (1980); 31, de Souza and Moreira (1994); 32, Galat *et al.* (1988); 33, Doregelo (1974).

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